

07/24/98  
1c533 U.S. PTO

Please type a plus sign (+) inside this box → ☐

PTO/SB/05 (4/98)  
Approved for use through 09/30/2000. OMB 0651-0032  
Patent and Trademark Office: U.S. DEPARTMENT OF COMMERCE

Under the Paperwork Reduction Act of 1995, no persons are required to respond to a collection of information unless it displays a valid OMB control number.

<b>UTILITY PATENT APPLICATION TRANSMITTAL</b> <small>(Only for new nonprovisional applications under 37 C.F.R. § 1.53(b))</small>	Attorney Docket No.	CVS-1
	First Inventor or Application Identifier	Siev
	Title	RESIN DERIVATIZATION METHOD AND USES THEREFOR
	Express Mail Label No.	EE470986382US

<b>APPLICATION ELEMENTS</b> <small>See MPEP chapter 600 concerning utility patent application contents.</small>	<b>ADDRESS TO:</b> Assistant Commissioner for Patents Box Patent Application Washington, DC 20231
1. <input checked="" type="checkbox"/> * Fee Transmittal Form (e.g., PTO/SB/17) <small>(Submit an original and a duplicate for fee processing)</small>	5. <input type="checkbox"/> Microfiche Computer Program (Appendix)
2. <input checked="" type="checkbox"/> Specification [Total Pages 90] - Descriptive title of the Invention - Cross References to Related Applications - Statement Regarding Fed sponsored R & D - Reference to Microfiche Appendix - Background of the Invention - Brief Summary of the Invention - Brief Description of the Drawings (if filed) - Detailed Description - Claim(s) - Abstract of the Disclosure	6. Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary) a. <input type="checkbox"/> Computer Readable Copy b. <input type="checkbox"/> Paper Copy (identical to computer copy) c. <input type="checkbox"/> Statement verifying identity of above copies
3. <input type="checkbox"/> Drawing(s) (35 U.S.C. 113) [Total Sheets ]	<b>ACCOMPANYING APPLICATION PARTS</b> 7. <input type="checkbox"/> Assignment Papers (cover sheet & document(s)) 8. <input type="checkbox"/> 37 C.F.R. § 3.73(b) Statement <input type="checkbox"/> Power of Attorney (when there is an assignee) 9. <input type="checkbox"/> English Translation Document (if applicable) 10. <input type="checkbox"/> Information Disclosure Statement (IDS)/PTO-1449 <input type="checkbox"/> Copies of IDS Citations 11. <input type="checkbox"/> Preliminary Amendment 12. <input checked="" type="checkbox"/> Return Receipt Postcard (MPEP 503) (Should be specifically itemized) * Small Entity <input type="checkbox"/> Statement filed in prior application, Status still proper and desired 13. <input type="checkbox"/> Statement(s) (PTO/SB/09-12) 14. <input type="checkbox"/> Certified Copy of Priority Document(s) (if foreign priority is claimed) 15. <input type="checkbox"/> Other: _____
4. Oath or Declaration [Total Pages 2] a. <input type="checkbox"/> Newly executed (original or copy) b. <input type="checkbox"/> Copy from a prior application (37 C.F.R. § 1.63(d)) (for continuation/divisional with Box 16 completed) i. <input type="checkbox"/> DELETION OF INVENTOR(S) Signed statement attached deleting inventor(s) named in the prior application, see 37 C.F.R. §§ 1.63(d)(2) and 1.33(b).	

\* NOTE FOR ITEMS 1 & 13: IN ORDER TO BE ENTITLED TO PAY SMALL ENTITY FEES, A SMALL ENTITY STATEMENT IS REQUIRED (37 C.F.R. § 1.27), EXCEPT IF ONE FILED IN A PRIOR APPLICATION IS RELIED UPON (37 C.F.R. § 1.28).

16. If a CONTINUING APPLICATION, check appropriate box, and supply the requisite information below and in a preliminary amendment:  
☐ Continuation ☐ Divisional ☐ Continuation-in-part (CIP) of prior application No: \_\_\_\_\_  
Prior application information: Examiner \_\_\_\_\_ Group / Art Unit: \_\_\_\_\_  
For CONTINUATION or DIVISIONAL APPS only: The entire disclosure of the prior application, from which an oath or declaration is supplied under Box 4b, is considered a part of the disclosure of the accompanying continuation or divisional application and is hereby incorporated by reference. The incorporation can only be relied upon when a portion has been inadvertently omitted from the submitted application parts.

**17. CORRESPONDENCE ADDRESS**

☐ Customer Number or Bar Code Label (Insert Customer No. or Attach bar code label here) or ☒ Correspondence address below

Name	Gerard H. Bencen, Esq. GERARD H. BENCEN, P.A.				
Address	426 Anderson Court				
City	Orlando	State	Florida	Zip Code	32801
Country	USA	Telephone	407-246-0444	Fax	407-246-1879

Name (Print/Type)	Gerard H. Bencen	Registration No. (Attorney/Agent)	35,746
Signature	<i>Gerard H. Bencen</i>	Date	July 24, 1998

Burden Hour Statement: This form is estimated to take 0.2 hours to complete. Time will vary depending upon the needs of the individual case. Any comments on the amount of time you are required to complete this form should be sent to the Chief Information Officer, Patent and Trademark Office, Washington, DC 20231. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Assistant Commissioner for Patents, Box Patent Application, Washington, DC 20231.

<h1 style="margin: 0;">FEE TRANSMITTAL</h1> <p style="font-size: small; margin: 5px 0;">Patent fees are subject to annual revision on October 1. These are the fees effective October 1, 1997.</p> <p style="font-size: x-small; margin: 0;">Small Entity payments <u>must</u> be supported by a small entity statement, otherwise large entity fees must be paid. See Forms PTO/SB/09-12.</p>		<b>Complete if Known</b>	
		Application Number	
		Filing Date	July 24, 1998
		First Named Inventor	Siev, Daniel V.
		Examiner Name	
		Group / Art Unit	
TOTAL AMOUNT OF PAYMENT		(\$) <u>3,230.00</u>	
		Attorney Docket No.	CVS-1

<p><b>METHOD OF PAYMENT (check one)</b></p> <p>1. <input type="checkbox"/> The Commissioner is hereby authorized to charge indicated fees and credit any over payments to:</p> <p>Deposit Account Number <span style="border: 1px solid black; display: inline-block; width: 150px; height: 1.2em; vertical-align: middle;"></span></p> <p>Deposit Account Name <span style="border: 1px solid black; display: inline-block; width: 150px; height: 1.2em; vertical-align: middle;"></span></p> <p><input type="checkbox"/> Charge Any Additional Fee Required Under 37 CFR 1.16 and 1.17      <input type="checkbox"/> Charge the Issue Fee Set in 37 CFR 1.18 at the Mailing of the Notice of Allowance</p> <p>2. <input checked="" type="checkbox"/> <b>Payment Enclosed:</b></p> <p style="margin-left: 20px;"><input checked="" type="checkbox"/> Check    <input type="checkbox"/> Money Order    <input type="checkbox"/> Other</p> <hr/> <p style="text-align: center;"><b>FEE CALCULATION</b></p> <p><b>1. BASIC FILING FEE</b></p> <table style="width: 100%; font-size: x-small;"> <thead> <tr> <th>Large Entity Fee Code (\$)</th> <th>Small Entity Fee Code (\$)</th> <th>Fee Description</th> <th>Fee Paid</th> </tr> </thead> <tbody> <tr> <td>101 790</td> <td>201 395</td> <td>Utility filing fee</td> <td style="border: 1px solid black; text-align: center;">790</td> </tr> <tr> <td>106 330</td> <td>206 165</td> <td>Design filing fee</td> <td style="border: 1px solid black;"></td> </tr> <tr> <td>107 540</td> <td>207 270</td> <td>Plant filing fee</td> <td style="border: 1px solid black;"></td> </tr> <tr> <td>108 790</td> <td>208 395</td> <td>Reissue filing fee</td> <td style="border: 1px solid black;"></td> </tr> <tr> <td>114 150</td> <td>214 75</td> <td>Provisional filing fee</td> <td style="border: 1px solid black;"></td> </tr> <tr> <td colspan="3" style="text-align: right;"><b>SUBTOTAL (1)</b></td> <td style="border: 1px solid black; text-align: center;">(\$)<u>790.00</u></td> </tr> </tbody> </table> <p><b>2. EXTRA CLAIM FEES</b></p> <table style="width: 100%; font-size: x-small;"> <thead> <tr> <th>Total Claims</th> <th>Extra Claims</th> <th>Fee from below</th> <th>Fee Paid</th> </tr> </thead> <tbody> <tr> <td>116</td> <td>20** = 96</td> <td>22</td> <td style="border: 1px solid black; text-align: center;">2,112</td> </tr> <tr> <td>7</td> <td>3** = 4</td> <td>82</td> <td style="border: 1px solid black; text-align: center;">328</td> </tr> <tr> <td colspan="3"></td> <td style="border: 1px solid black;"></td> </tr> </tbody> </table> <p><small>**or number previously paid, if greater; For Reissues, see below</small></p> <table style="width: 100%; font-size: x-small;"> <thead> <tr> <th>Large Entity Fee Code (\$)</th> <th>Small Entity Fee Code (\$)</th> <th>Fee Description</th> <th>Fee Paid</th> </tr> </thead> <tbody> <tr> <td>103 22</td> <td>203 11</td> <td>Claims in excess of 20</td> <td style="border: 1px solid black;"></td> </tr> <tr> <td>102 82</td> <td>202 41</td> <td>Independent claims in excess of 3</td> <td style="border: 1px solid black;"></td> </tr> <tr> <td>104 270</td> <td>204 135</td> <td>Multiple dependent claim, if not paid</td> <td style="border: 1px solid black;"></td> </tr> <tr> <td>109 82</td> <td>209 41</td> <td>** Reissue independent claims over original patent</td> <td style="border: 1px solid black;"></td> </tr> <tr> <td>110 22</td> <td>210 11</td> <td>** Reissue claims in excess of 20 and over original patent</td> <td style="border: 1px solid black;"></td> </tr> <tr> <td colspan="3" style="text-align: right;"><b>SUBTOTAL (2)</b></td> <td style="border: 1px solid black; text-align: center;">(\$)<u>2,440.00</u></td> </tr> </tbody> </table>	Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description	Fee Paid	101 790	201 395	Utility filing fee	790	106 330	206 165	Design filing fee		107 540	207 270	Plant filing fee		108 790	208 395	Reissue filing fee		114 150	214 75	Provisional filing fee		<b>SUBTOTAL (1)</b>			(\$) <u>790.00</u>	Total Claims	Extra Claims	Fee from below	Fee Paid	116	20** = 96	22	2,112	7	3** = 4	82	328					Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description	Fee Paid	103 22	203 11	Claims in excess of 20		102 82	202 41	Independent claims in excess of 3		104 270	204 135	Multiple dependent claim, if not paid		109 82	209 41	** Reissue independent claims over original patent		110 22	210 11	** Reissue claims in excess of 20 and over original patent		<b>SUBTOTAL (2)</b>			(\$) <u>2,440.00</u>	<p><b>3. ADDITIONAL FEES</b></p> <table style="width: 100%; font-size: x-small;"> <thead> <tr> <th>Large Entity Fee Code (\$)</th> <th>Small Entity Fee Code (\$)</th> <th>Fee Description</th> <th>Fee Paid</th> </tr> </thead> <tbody> <tr><td>105 130</td><td>205 65</td><td>Surcharge - late filing fee or oath</td><td style="border: 1px solid black;"></td></tr> <tr><td>127 50</td><td>227 25</td><td>Surcharge - late provisional filing fee or cover sheet</td><td style="border: 1px solid black;"></td></tr> <tr><td>139 130</td><td>139 130</td><td>Non-English specification</td><td style="border: 1px solid black;"></td></tr> <tr><td>147 2,520</td><td>147 2,520</td><td>For filing a request for reexamination</td><td style="border: 1px solid black;"></td></tr> <tr><td>112 920*</td><td>112 920*</td><td>Requesting publication of SIR prior to Examiner action</td><td style="border: 1px solid black;"></td></tr> <tr><td>113 1,840*</td><td>113 1,840*</td><td>Requesting publication of SIR after Examiner action</td><td style="border: 1px solid black;"></td></tr> <tr><td>115 110</td><td>215 55</td><td>Extension for reply within first month</td><td style="border: 1px solid black;"></td></tr> <tr><td>116 400</td><td>216 200</td><td>Extension for reply within second month</td><td style="border: 1px solid black;"></td></tr> <tr><td>117 950</td><td>217 475</td><td>Extension for reply within third month</td><td style="border: 1px solid black;"></td></tr> <tr><td>118 1,510</td><td>218 755</td><td>Extension for reply within fourth month</td><td style="border: 1px solid black;"></td></tr> <tr><td>128 2,060</td><td>228 1,030</td><td>Extension for reply within fifth month</td><td style="border: 1px solid black;"></td></tr> <tr><td>119 310</td><td>219 155</td><td>Notice of Appeal</td><td style="border: 1px solid black;"></td></tr> <tr><td>120 310</td><td>220 155</td><td>Filing a brief in support of an appeal</td><td style="border: 1px solid black;"></td></tr> <tr><td>121 270</td><td>221 135</td><td>Request for oral hearing</td><td style="border: 1px solid black;"></td></tr> <tr><td>138 1,510</td><td>138 1,510</td><td>Petition to institute a public use proceeding</td><td style="border: 1px solid black;"></td></tr> <tr><td>140 110</td><td>240 55</td><td>Petition to revive - unavoidable</td><td style="border: 1px solid black;"></td></tr> <tr><td>141 1,320</td><td>241 660</td><td>Petition to revive - unintentional</td><td style="border: 1px solid black;"></td></tr> <tr><td>142 1,320</td><td>242 660</td><td>Utility issue fee (or reissue)</td><td style="border: 1px solid black;"></td></tr> <tr><td>143 450</td><td>243 225</td><td>Design issue fee</td><td style="border: 1px solid black;"></td></tr> <tr><td>144 670</td><td>244 335</td><td>Plant issue fee</td><td style="border: 1px solid black;"></td></tr> <tr><td>122 130</td><td>122 130</td><td>Petitions to the Commissioner</td><td style="border: 1px solid black;"></td></tr> <tr><td>123 50</td><td>123 50</td><td>Petitions related to provisional applications</td><td style="border: 1px solid black;"></td></tr> <tr><td>126 240</td><td>126 240</td><td>Submission of Information Disclosure Stmt</td><td style="border: 1px solid black;"></td></tr> <tr><td>581 40</td><td>581 40</td><td>Recording each patent assignment per property (times number of properties)</td><td style="border: 1px solid black;"></td></tr> <tr><td>146 790</td><td>246 395</td><td>Filing a submission after final rejection (37 CFR 1.129(a))</td><td style="border: 1px solid black;"></td></tr> <tr><td>149 790</td><td>249 395</td><td>For each additional invention to be examined (37 CFR 1.129(b))</td><td style="border: 1px solid black;"></td></tr> <tr><td colspan="3">Other fee (specify) _____</td><td style="border: 1px solid black;"></td></tr> <tr><td colspan="3">Other fee (specify) _____</td><td style="border: 1px solid black;"></td></tr> <tr> <td colspan="3" style="text-align: right;"><b>SUBTOTAL (3)</b></td> <td style="border: 1px solid black; text-align: center;">(\$)<u>0</u></td> </tr> </tbody> </table> <p><small>* Reduced by Basic Filing Fee Paid</small></p>	Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description	Fee Paid	105 130	205 65	Surcharge - late filing fee or oath		127 50	227 25	Surcharge - late provisional filing fee or cover sheet		139 130	139 130	Non-English specification		147 2,520	147 2,520	For filing a request for reexamination		112 920*	112 920*	Requesting publication of SIR prior to Examiner action		113 1,840*	113 1,840*	Requesting publication of SIR after Examiner action		115 110	215 55	Extension for reply within first month		116 400	216 200	Extension for reply within second month		117 950	217 475	Extension for reply within third month		118 1,510	218 755	Extension for reply within fourth month		128 2,060	228 1,030	Extension for reply within fifth month		119 310	219 155	Notice of Appeal		120 310	220 155	Filing a brief in support of an appeal		121 270	221 135	Request for oral hearing		138 1,510	138 1,510	Petition to institute a public use proceeding		140 110	240 55	Petition to revive - unavoidable		141 1,320	241 660	Petition to revive - unintentional		142 1,320	242 660	Utility issue fee (or reissue)		143 450	243 225	Design issue fee		144 670	244 335	Plant issue fee		122 130	122 130	Petitions to the Commissioner		123 50	123 50	Petitions related to provisional applications		126 240	126 240	Submission of Information Disclosure Stmt		581 40	581 40	Recording each patent assignment per property (times number of properties)		146 790	246 395	Filing a submission after final rejection (37 CFR 1.129(a))		149 790	249 395	For each additional invention to be examined (37 CFR 1.129(b))		Other fee (specify) _____				Other fee (specify) _____				<b>SUBTOTAL (3)</b>			(\$) <u>0</u>
Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description	Fee Paid																																																																																																																																																																																														
101 790	201 395	Utility filing fee	790																																																																																																																																																																																														
106 330	206 165	Design filing fee																																																																																																																																																																																															
107 540	207 270	Plant filing fee																																																																																																																																																																																															
108 790	208 395	Reissue filing fee																																																																																																																																																																																															
114 150	214 75	Provisional filing fee																																																																																																																																																																																															
<b>SUBTOTAL (1)</b>			(\$) <u>790.00</u>																																																																																																																																																																																														
Total Claims	Extra Claims	Fee from below	Fee Paid																																																																																																																																																																																														
116	20** = 96	22	2,112																																																																																																																																																																																														
7	3** = 4	82	328																																																																																																																																																																																														
Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description	Fee Paid																																																																																																																																																																																														
103 22	203 11	Claims in excess of 20																																																																																																																																																																																															
102 82	202 41	Independent claims in excess of 3																																																																																																																																																																																															
104 270	204 135	Multiple dependent claim, if not paid																																																																																																																																																																																															
109 82	209 41	** Reissue independent claims over original patent																																																																																																																																																																																															
110 22	210 11	** Reissue claims in excess of 20 and over original patent																																																																																																																																																																																															
<b>SUBTOTAL (2)</b>			(\$) <u>2,440.00</u>																																																																																																																																																																																														
Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description	Fee Paid																																																																																																																																																																																														
105 130	205 65	Surcharge - late filing fee or oath																																																																																																																																																																																															
127 50	227 25	Surcharge - late provisional filing fee or cover sheet																																																																																																																																																																																															
139 130	139 130	Non-English specification																																																																																																																																																																																															
147 2,520	147 2,520	For filing a request for reexamination																																																																																																																																																																																															
112 920*	112 920*	Requesting publication of SIR prior to Examiner action																																																																																																																																																																																															
113 1,840*	113 1,840*	Requesting publication of SIR after Examiner action																																																																																																																																																																																															
115 110	215 55	Extension for reply within first month																																																																																																																																																																																															
116 400	216 200	Extension for reply within second month																																																																																																																																																																																															
117 950	217 475	Extension for reply within third month																																																																																																																																																																																															
118 1,510	218 755	Extension for reply within fourth month																																																																																																																																																																																															
128 2,060	228 1,030	Extension for reply within fifth month																																																																																																																																																																																															
119 310	219 155	Notice of Appeal																																																																																																																																																																																															
120 310	220 155	Filing a brief in support of an appeal																																																																																																																																																																																															
121 270	221 135	Request for oral hearing																																																																																																																																																																																															
138 1,510	138 1,510	Petition to institute a public use proceeding																																																																																																																																																																																															
140 110	240 55	Petition to revive - unavoidable																																																																																																																																																																																															
141 1,320	241 660	Petition to revive - unintentional																																																																																																																																																																																															
142 1,320	242 660	Utility issue fee (or reissue)																																																																																																																																																																																															
143 450	243 225	Design issue fee																																																																																																																																																																																															
144 670	244 335	Plant issue fee																																																																																																																																																																																															
122 130	122 130	Petitions to the Commissioner																																																																																																																																																																																															
123 50	123 50	Petitions related to provisional applications																																																																																																																																																																																															
126 240	126 240	Submission of Information Disclosure Stmt																																																																																																																																																																																															
581 40	581 40	Recording each patent assignment per property (times number of properties)																																																																																																																																																																																															
146 790	246 395	Filing a submission after final rejection (37 CFR 1.129(a))																																																																																																																																																																																															
149 790	249 395	For each additional invention to be examined (37 CFR 1.129(b))																																																																																																																																																																																															
Other fee (specify) _____																																																																																																																																																																																																	
Other fee (specify) _____																																																																																																																																																																																																	
<b>SUBTOTAL (3)</b>			(\$) <u>0</u>																																																																																																																																																																																														

<b>SUBMITTED BY</b>				<b>Complete (if applicable)</b>	
Typed or Printed Name	Gerard H. Bencen, Esq.			Reg. Number	35,746
Signature			Date	7/24/98	Deposit Account User ID

Burden Hour Statement: This form is estimated to take 0.2 hours to complete. Time will vary depending upon the needs of the individual case. Any comments on the amount of time you are required to complete this form should be sent to the Chief Information Officer, Patent and Trademark Office, Washington, DC 20231. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Assistant Commissioner for Patents, Washington, DC 20231.

**TITLE OF THE INVENTION****RESIN DERIVATIZATION METHOD AND USES THEREOF****BACKGROUND OF THE INVENTION**

5

**1. FIELD OF THE INVENTION:**

This invention provides processes and methods for making derivatized resins useful in the arts of solid-phase peptide synthesis, combinatorial chemistry, peptide or protein  
10 purification and separation, and other arts. Resins prepared according to the method of this invention are solid support reagents suitable for use in conventional automated, semi-automated or manual peptide synthesis using protected amino acids or amino acid analogs, to give a protected peptide (or peptide analog) aldehyde or ketoamide, attached to the support reagent. The product peptide aldehyde or ketoamide peptide or peptide  
15 analog is cleaved from the support and deprotected to give the desired peptide or peptide analog in good yield. As opposed to known methods for peptide aldehyde or peptide ketoamide synthesis, the process of the present invention provides, among other benefits, a method for solid-phase peptide or peptide analog synthesis that minimizes the amount of solution-phase synthetic steps required.

20

**2. BACKGROUND:**

In the field of protease inhibition, a number of peptide and peptide analog inhibitors have been developed, as have methods for their synthesis (see U.S. Patent Nos. 5,283,293;  
25 5,367,072; 5,371,072; 5,492,895; 5,514,777; 5,597,804; 5,637,599; 5,646,165; 5,656,600; 5,656,645; 5,681,844; 5,696,231; 5,703,208; 5,714,580; 5,731,413; 5,739,112; all of which are herein incorporated by reference. Accordingly, there have been significant developments in the field of protease inhibition and the development of structure-activity profiles for a number of classes of protease inhibitors.

Of particular significance to the field of protease inhibition is the development of synthetic methods of ever increasing efficiency. Accordingly, there is a continued need in the art for efficient processes for solid-phase production of protease inhibitors and other peptidyl and peptidomimetic compounds. The present invention provides processes that  
5 are an improvement on existing solid-phase synthetic methods and resins.

In the art of protease inhibition, compounds including peptides, peptide analogs, and peptidomimetic compounds having an aldehyde group on the C-terminus of the peptide have been found to be good transition-state inhibitors, many of which show marked  
10 selectivity for specific proteolytic enzymes. However, significant difficulty has been encountered in the synthetic production of such molecules. Several methods for solution-phase synthesis of such compounds have been reported (see, for Example, U.S. Patent No. 5,514,777 and references cited therein).

15 Methods for the solution-phase synthesis of peptide aldehydes have been reported. (McConnell, R. M. et al., J. Med. Chem. 33:86-93, 1990, and references cited therein; see also Kawamura et al., Chem. Pharm. Bull. 17:1902, 1969; Someno et al., Chem. Pharm. Bull. 34, 1748, 1986). The use of semicarbazides as aldehyde protecting reagents for the solution synthesis of peptide aldehydes has also been reported. (Westerik and Wolfinden,  
20 J. Biol. Chem., 247, 8195 (1972); Ito et al., Chem. Pharm. Bull. 23, 3081 (1975)). The use of soluble semicarbazide functionalized polymer has been reported for the manual preparation of some peptide aldehydes. (Galpin, et al., Pept. Struct. Funct. Proc. Am. Pept. Symp., 9th, 799-802, 1985, Edited by: Deber, C. M., Hruby, V. J., Kopple, K.D., Pierce Chem. Co., Rockford, Ill.). However such supports were reported not to be  
25 suitable for the automatic synthesis of peptide aldehydes, since they dissolve in the solvents used for the coupling steps.

The use of semicarbazone intermediates has been reported in the synthesis of peptidyl argininals. The unsubstituted semicarbazone, N<sup>g</sup>-nitro-L-argininal semicarbazone was  
30 used as an intermediate in the synthesis of peptidyl argininals. McConnell, R. M., et al.,

J. Med. Chem., 33:86 (1990); R. M. McConnel, J.L. York, D. Frissell, C. Ezell, J. Med. Chem. 36:1084-1089 (1993). N<sup>ε</sup>-nitro-L-argininal semicarbazonyl-4-methylcyclohexane carboxylic acid was reported as an intermediate in the preparation of peptide aldehydes by a solid phase method. Murphy, A. M. et al., J. Am. Chem. Soc., 114:3156 (1992); and  
5 Webb, T. R., U.S. Patent No. 5,283,293 (Feb. 1, 1994), and U.S. Patent No. 5,367,072 (Nov. 22, 1994). N<sup>ε</sup>-nitro-L-argininal semicarbazonyl-4-diphenylmethane was reported as an intermediate for the solution-phase synthesis of peptidyl argininals. Brunck, T. K. et al., WO 93/14779 (1993). Solution and solid phase syntheses of peptidyl argininals was disclosed in U.S. Patent No. 5,731,413 (Mar. 24, 1998). The disclosed methods  
10 implicated the use of an intermediate arginine cyclol N- or O-linked to a polymeric support.

The present invention provides a new method for the efficient production of argininal and other peptide or peptidomimetic aldehydes and ketoamides by solid phase synthesis,  
15 wherein a linkage between the aldehyde or ketoamide and a solid support provides for simplified chemistry with high yields of the desired product aldehyde or ketoamide. As opposed to known methods for peptide aldehyde synthesis, the process of the present invention provides a method for solid-phase peptide or peptide analog synthesis which provides the option of reduced reliance on solution-phase synthetic manipulation to  
20 produce desired aldehyde product. Accordingly, those skilled in the art will appreciate that the method of this invention represents a substantial contribution to the art in that solution-phase reactions, which may result in reduced product yield and which may require days if not weeks to accomplish, are accomplished according to the method of this invention within hours, due to reduced solution-phase synthetic chemistry heretofore  
25 required. By substantially reducing the product development cycle, true solid-phase combinatorial chemistry is enabled, including for heretofore difficult to synthesize, pharmacologically significant, aldehyde and ketoamide protease inhibitors.

## SUMMARY OF THE INVENTION

Methods for synthesizing a derivatized solid support, the prototypical example of which is hydrazyl-carbonyl-amino methylated polystyrene (also referred to herein as “HCAM” resin or solid support), are provided by this invention. The processes according to this invention are used for preparing a functionalized solid support having the formula (I):

la (I):

(I)  $R_4-NH-(C=X)-Y-Z-SS$  wherein:

$R_4$  is  $-NH-R_3$ ,  $-NH_2$ ,  $-OH$ , or  $-O-R_3$ , wherein  $R_3$  is a protecting group, provided that when  $R_4$  is  $-NH-R_3$  or  $-O-R_3$ , then the protecting group is removed and replaced by  $-H$  in the final product (I);

$X$  is O, S, or  $NR_7$ ;

$R_7$  is H, alkyl, alkenyl, aryl, aralkyl, cycloalkyl, or heterocycle;

$Y$  is absent,  $-NH-$ , or  $-CH_2-$ ;

$Z$  is absent or is a substituent selected from the group consisting of  $-NH-$ ,  $-O-$ ,  $-(C=O)-$ ,  $-S-$ ,  $SO_2-$ , alkyl, alkenyl, aryl, aralkyl, cycloalkyl, heterocycle, and combinations thereof, provided that when  $Y$  is absent and  $X$  is O or S,  $Z$  does not comprise  $-(C=O)-$  immediately adjacent to  $-(C=X)-$ , and when  $Y$  is  $-NH-$  and  $Z$  comprises an  $-NH-$  or an  $-S-$ , at least one carbon atom separates  $Y$  and the  $-NH-$  or  $-S-$  of  $Z$ , wherein, under conditions for peptide synthesis, functional groups of  $Z$  are protected; and

$SS$  is a solid support.

The derivatized solid support (I) is available for reaction at  $R_4$  with appropriately protected aldehyde or ketoamide moieties, extension of a peptide or peptide analog chain thereon, and subsequent cleavage thereof from the resin to yield free aldehyde or ketoamide product.

The product produced according to the methods of this invention is useful as a solid support for solid-phase peptide synthesis, combinatorial chemistry, the purification or isolation of peptides or proteins, and other uses.

5 Accordingly, it is one object of the present invention to provide a method whereby solid-phase peptide or peptidomimetic compounds comprising an aldehyde or ketoamide at the P1 position (i.e. the carboxy-terminal end) of the peptide or peptidomimetic compound is achieved, with reduced need for solution-phase synthetic manipulation.

10 A further object of this invention is to provide a method for derivatization of solid supports for use in solid-phase peptide synthesis.

A further object of this invention is to provide methods for derivatization of solid supports for use in combinatorial chemistry.

15

A further object of this method is to provide procedures for derivatization of solid supports for use in the purification or isolation of peptides or proteins.

Additional objects of this invention will become apparent from a review of the full  
20 disclosure, including the appended claims.

## **DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS**

### **1. Definitions:**

25

The present invention is directed to a method for production of derivatized solid supports, as well as methods of using the thus-derivatized solid supports in the arts of solid-phase peptide synthesis, combinatorial chemistry and peptide or protein purification and separation.

30

As used in this disclosure, the following terms have the defined meaning, unless expressly modified in the context in which the term is used:

SS is a polymeric solid support that is stable in the presence of acids, bases and/or other reagents; a “solid support” is any form of bead or resin typically used in the art of peptide synthesis to provide a “handle” whereby a growing synthetic peptide chain may be made available for synthetic manipulation without the risk of loss in peptide yield typically experienced when such syntheses are conducted in solution; the terms “solid support” and “resin” are used interchangeably. The term “solid support”, “SS” or “support” refer to a solid particulate, insoluble material to which a linker moiety of the present invention is linked and from which a peptide or peptide analog may be synthesized. Supports used in synthesizing peptides and peptide analogs are typically substantially inert and nonreactive with the reagents used in the synthesis of peptides and peptide analogs, particularly once an initial linkage between an aldehyde or ketoamide has been established (i.e., a P1 position has been established), according to the method of this invention. Preferably, a solid support is a cross-linked resin, such as polystyrene.

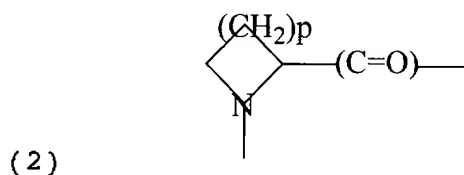
NH<sub>2</sub>-SS is a SS comprising at least one functional amino group available for formation of an amide (peptide) bond.

The term “amino acid” refers to natural amino acids or unnatural amino acids, and amino acid analogs in their D and L stereoisomers if their structure allows such stereoisomeric forms. Natural amino acids include alanine (Ala), arginine (Arg), asparagine (Asn), aspartic acid (Asp), cysteine (Cys), glutamine (Gln), glutamic acid (Glu), glycine (Gly), histidine (His), isoleucine (Ile), leucine (Leu), lysine (Lys), methionine (Met), phenylalanine (Phe), proline (Pro), serine (Ser), threonine (Thr), tryptophan (Trp), tyrosine (Tyr) and valine (Val). Unnatural amino acids include, but are not limited to azetidinecarboxylic acid, 2-aminoadipic acid, 3-aminoadipic acid, beta-alanine, aminopropionic acid, 2-aminobutyric acid, 4-aminobutyric acid, 6-aminocaproic acid, 2-aminoheptanoic acid, 2-aminoisobutyric acid, 3-aminoisobutyric acid, 2-aminopimelic



acid, 2,4-diaminoisobutyric acid, desmosine, 2,2'-diaminopimelic acid, 2,3-diaminopropionic acid, n-ethylglycine, N-ethylasparagine, hydroxylysine, all-hydroxylysine, 3-hydroxyproline, 4-hydroxyproline, isodesmosine, allo-isoleucine, N-methylglycine, N-methylisoleucine, N-methylvaline, norvaline, norleucine, ornithine and  
 5 pipecolic acid. Amino acid analogs include, but are not limited to, the natural and unnatural amino acids which are chemically blocked, reversibly or irreversibly, or modified on their N-terminal amino group or their side-chain groups, as for example, methionine sulfoxide, methionine sulfone, S-(carboxymethyl)-cysteine, S-(carboxymethyl)-cysteine sulfoxide and S-(carboxymethyl)-cysteine sulfone, aspartic  
 10 acid-(beta methyl ester), N-ethylglycine, and alanine carboxamide.

The term "amino acid residue" refers to radicals having the structure (1)  $\text{-NH-R-C(O)-}$ , wherein R typically is  $\text{-CH(R*)-}$ , wherein R\* is H or a carbon containing substituent; or



15 wherein p is 1, 2, or 3, representing the azetidinecarboxylic acid, proline or pipecolic acid residues, respectively.

A peptide or peptide analog is a molecule comprising at least two amino acids or amino acid analogs linked through peptide (amide) linkages.

20

A peptidomimetic compound is any compound which structurally resembles or mimics a natural peptidyl array, and compounds comprising such residues; compounds which, although not a natural peptide, in the sense that it either contains no amino acids or contains amino acid analogs, exhibits a biological activity of a known peptidyl  
 25 compound.

The terms "good leaving group" or "leaving group" are used herein to define a molecular substituent which, when used in conducting chemical syntheses, exhibits the desirable

properties of being labile under defined synthetic conditions, and of being easily separated from synthetic products under defined conditions. Examples of such leaving groups include, but are not limited to, hydrogen, hydroxyl radicals, halogen atoms, p-nitrophenoxide, water, methyl groups, and the like.

5

The term “protecting group” is used herein to refer to well known moieties which have the desirable property of preventing specific chemical reactions at a site on a molecule undergoing chemical modification intended to be left unaffected by the particular chemical modification, while at the same time being easily removed from the molecule under conditions that do not adversely affect other sites in the modified molecule. Those skilled in the art have a wide variety of known protecting groups to choose from, depending on the nature of the chemical site to be protected. Reference is made, for example, to “Protective Groups in Organic Synthesis”, T. Greene, (John Wiley & Sons, Inc., 1991), and to “Solid Phase Peptide Synthesis,” Stewart and Young (Pierce Chemical Co., 1984), herein incorporated by reference for this and other purposes. Examples of protecting groups known in the art include, but are not limited to, Cbz, Boc, Alloc, Fmoc, Troc, Teoc ( $\text{Me}_3\text{Si}(\text{CH}_2)_2\text{OCO}$ ), PMC, and the like, and others disclosed herein.

10

15

20

The term “alkyl” refers to saturated aliphatic groups including straight-chain, branched-chain and cyclic groups.

The term “alkenyl” refers to unsaturated aliphatic groups having at least one double bond.

25

The term “aryl” refers to aromatic groups which have at least one ring having a conjugated pi electron system and includes carbocyclic aryl, heterocyclic aryl and biaryl groups, all of which may optionally be substituted with a substituent selected from, but not limited to, lower alkyl of one to ten carbon atoms; alkenyl; nitro; cyano; halo;  $-\text{S}(\text{O})_q-$ , wherein, for purposes of this definition, q is 0, 1, or 2; carboxylic acid or carboxylic acid derivatives, esters amides, and the like.

30

The term “aralkyl” refers to an alkyl group substituted with an aryl group. Exemplary aralkyl groups include, but are not limited to, benzyl, picolyl, and the like, which may optionally be substituted with a substituent selected from, but not limited to, lower alkyl of one to ten carbon atoms; alkenyl; nitro; cyano; halo; -S(O)<sub>q</sub>-, wherein, for purposes of this definition, q is 0, 1, or 2; carboxylic acid or carboxylic acid derivatives, esters amides, and the like.

The term “cycloalkyl” refers to an alkyl in which at least a portion of the molecule is in a closed ring configuration. Exemplary cycloalkyl groups include, but are not limited to, cyclohexyl, cyclopropyl, cyclopentyl and cycloheptyl.

The term “heterocycle” refers to any compound having a closed cyclic structure in which at least one atom thereof is other than a carbon atom. For example, cyclic alkyl, cyclic aryl, cyclic aralkyl compounds containing a nitrogen, oxygen or sulfur atom in the cyclic structure are heterocycles.

The term “halo” or “halogen” refers to fluorine, chlorine, bromine and iodine.

The term “non-adverse conditions” describes conditions of reaction or synthesis which do not substantially adversely affect the skeleton of the peptide analog and/or its amino acid (and/or amino acid analog) components. One skilled in the art can readily identify functionalities, coupling procedures, deprotection procedures and cleavage conditions which meet these criteria.

A semicarbazone is a derivative formed by the reaction of a semicarbazide with an aldehyde or ketone of the general formula R'(R')C=NNH(CO)NHR'R', wherein for each occurrence of R', the R' is independently selected to be one of H, alkyl, aryl, and other organic residues, such as, for purposes of this invention, a solid support, an amino acid, a peptide or peptide analog.

A semicarbazide is a reagent of the formula  $H_2NNH(CO)NH_2$  or a derivative thereof of formula  $H_2NNH(CO)NR'R'$ , which is known to react with a variety of electrophiles, including carbonyl compounds such as aldehydes and ketones, wherein for each occurrence of  $R'$ , the  $R'$  is independently selected to be one of H, alkyl, aryl, and other organic residues, such as, for purposes of this invention, a solid support, an amino acid, a peptide or peptide analog.

Alloc is allyloxycarbonyl

Alloc-Cl is allyl chloroformate

10 A.M. resin is amino-methylated polystyrene resin

Boc is tert-butyloxy carbonyl or tert-butoxycarbonyl

Bom is benzyloxymethyl

BOP is benzotriazole-yl-oxy-tris-(dimethylamino)phosphonium hexafluorophosphate

Cbz is benzyloxycarbonyl or carbobenzyloxy

15 2-Clz is 2-chlorobenzyloxycarbonyl

DCM is dichloromethane

DIEA is N, N-diisopropylethylamine

DMF is N, N-dimethylformamide

DMSO is dimethylsulfoxide

20 EDC is 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride salt

Fmoc is 9-fluorenylmethyloxycarbonyl

HCAM is hydrazyl-carbonyl-amino methylated polystyrene

HF is hydrogen fluoride

HOBt or HOBt is 1-hydroxybenzotriazole monohydrate

25 HPLC is high pressure liquid chromatography; high performance liquid chromatography

LAH is lithium aluminum hydride

MS is mass spectrometry

Mts is mesitylene-2-sulphonyl

NMM is N-methylmorpholine (also referred to as 4-methylmorpholine)

30 NMR is nuclear magnetic resonance spectroscopy

PG is protecting group

PMC is 2,2,5,7,8-pentamethylchroman-6-sulfonyl

PyBOP is Benzotriazole-yl-oxy-tris-pyrrolidino-phosphonium hexafluorophosphate

(Ph<sub>3</sub>P)<sub>4</sub>Pd is tetrakis-(triphenylphosphine)palladium(0)

5 r.t. or RT is room temperature

TBTU is 2-(1H-benzotriazole-1-yl)-1,1,3,3-tetramethyluronium tetrafluoroborate

TeOC is Me<sub>3</sub>Si(CH<sub>2</sub>)OCO

TFA is trifluoroacetic acid

THF is tetrahydrofuran

10 TLC or tlc is thin layer chromatography

TFMSA is trifluoromethylsulfonic acid, also commonly referred to as “triflic acid”

TMSOTf is trimethylsilyltrifluoro methane sulfonate

Tos is p-toluenesulfonyl, also referred to as Tosyl or Ts

Troc is trichloroethoxycarbonyl (an amine protecting group removable with zinc)

15 NMR designations:

s is singlet

d is doublet

m is multiplet

br is broad peak

20 t is triplet

q is quartet

## **2. Description of the Preferred Embodiments:**

25 According to the present invention, synthetic methods are provided for production of derivatized resin, also referred to as a derivatized solid support, and production of peptide or peptidomimetic products using those resins. From the present disclosure, it will be apparent that the methods disclosed herein are useful for the synthesis of peptides, peptide analogs, and peptidomimetics, and while all of these possibilities may not be

recited in a particular context, production of all of these possibilities is intended. The method according to this invention comprises the steps of:

**A. Preparing a derivatized resin optimized for reaction with appropriately protected aldehyde or ketoamide moieties, wherein said resin is represented by the formula (I):**

(I)  $R_4-NH-(C=X)-Y-Z-SS$  wherein:

$R_4$  is  $-NH-R_3$ ,  $-NH_2$ ,  $-OH$ , or  $-O-R_3$ , wherein  $R_3$  is a protecting group, provided that when  $R_4$  is  $-NH-R_3$  or  $-O-R_3$ , then the protecting group is removed and replaced by  $-H$  in the final product (I);

$X$  is  $O$ ,  $S$ , or  $NR_7$ ;

$R_7$  is  $H$ , alkyl, alkenyl, aryl, aralkyl, cycloalkyl, or heterocycle;

$Y$  is absent,  $-NH-$ , or  $-CH_2-$ ;

$Z$  is absent or is a substituent selected from the group consisting of  $-NH-$ ,  $-O-$ ,  $-(C=O)-$ ,  $-S-$ ,  $SO_2-$ , alkyl, alkenyl, aryl, aralkyl, cycloalkyl, heterocycle, and combinations thereof, provided that when  $Y$  is absent and  $X$  is  $O$  or  $S$ ,  $Z$  does not comprise  $-(C=O)-$  immediately adjacent to  $-(C=X)-$ , and when  $Y$  is  $-NH-$  and  $Z$  comprises an  $-NH-$  or an  $-S-$ , at least one carbon atom separates  $Y$  and the  $-NH-$  or  $-S-$  of  $Z$ , wherein, under conditions for peptide synthesis, functional groups of  $Z$  are protected;

$SS$  is a solid support; and

**B. Reacting an aldehyde or ketoamide with the derivatized resin of step (A), thereby producing an immobilized aldehyde or ketoamide.**

It will be appreciated that, by virtue of immobilization, the aldehyde or ketoamide may be purified or isolated from a solution and, optionally, subsequently released as the free aldehyde or ketoamide. Alternatively, solid-phase chemical modification of said

immobilized aldehyde or ketoamide, including but not limited to solid-phase peptide synthesis thereon or combinatorial chemistry, may be conducted. Accordingly, the process of this invention may further comprise the additional steps of:

- 5     **C.     Optionally performing solid-phase chemistry on the immobilized aldehyde or ketoamide of step (B), selected from peptide synthesis, synthesis of peptide analogs, and combinatorial chemistry, thereby producing a modified aldehyde or ketoamide.**

10     Finally, the free aldehyde or ketoamide, modified or not, may be released from the resin by cleavage therefrom. Therefore, the method of this invention may further comprise the step of:

- 15     **D.     Cleaving, deprotecting and recovering the aldehyde or ketoamide of step (B) or the modified aldehyde or ketoamide of step (C) as the free aldehyde or ketoamide.**

These aspects of the invented process are described in greater detail below:

- 20     **A.     Preparing a derivatized resin (I) optimized for reaction with appropriately protected aldehyde or ketoamide moieties:**

- a.     The Derivatized Resin Produced According to the Methods of this Invention:

25     The invention according to the present disclosure comprises the production of derivatized resins for use in any of a number of different methods, including solid-phase peptide synthesis, combinatorial chemistry and peptide or protein purification or isolation.

30     For reaction with appropriately protected aldehyde or ketoamide moieties, the methods of this aspect of the invention produce a functionalized solid support represented by formula (I):

(I) R4-NH-(C=X)-Y-Z-SS wherein:

R4 is -NH-R3, -NH<sub>2</sub>, -OH, or -O-R3, wherein R3 is a protecting group, provided that when R4 is -NH-R3 or -O-R3, then the protecting group is removed and replaced by -H in the final product

(I);

X is O, S, or NR7;

R7 is H, alkyl, alkenyl, aryl, aralkyl, cycloalkyl, or heterocycle;

Y is absent, -NH-, or -CH<sub>2</sub>-;

Z is absent or is a substituent selected from the group consisting of -NH-, -O-, -(C=O)-, -S-, SO<sub>2</sub>-, alkyl, alkenyl, aryl, aralkyl, cycloalkyl, heterocycle, and combinations thereof, provided that when Y is absent and X is O or S, Z does not comprise -(C=O)- immediately adjacent to -(C=X)-, and when Y is -NH- and Z comprises an -NH- or an -S-, at least one carbon atom separates Y and the -NH- or -S- of Z, wherein, under conditions for peptide synthesis, functional groups of Z are protected;

SS is a solid support.

b. Reactants and Synthetic Routes for Production of the Derivatized Resin of Formula (I):

The solid support of formula (I) is prepared by any of the following synthetic routes, wherein reactants as defined below are utilized. Each of these reactants is either readily available from commercial sources or may be prepared by those of ordinary skill in the art using standard synthetic methods, coupled with the disclosure and guidance provided herein. The reactants utilized have the following formulae and substituent definitions, wherein the reactant name or definition associated with a given reactant may be used to refer both to the generic formula thereof as well as to specific embodiments thereof. Alternatively, in certain instances, a species of a generic reactant may be given an new designation for clarity. In the following reactant definitions, variables defined above in



the definition of the derivatized resin (I) have the same definitions below, unless otherwise specified:

- (A) R-Y-Z-SS, wherein R is a leaving group.
- 5 (B) R1-(C=X)-R2, wherein R1 and R2 are independently selected leaving groups, same or different.
- (C) R1-(C=X)-Y-Z-SS, wherein all variables are as defined above. This reactant (C) includes but is not limited to a commercially available carboxylated resin of formula:
- 10 HOOC-Y-Z-SS, and is prepared by carboxylating a commercially available resin bearing at least one functional amino group according to methods known in the art.
- (D) R4-NH<sub>2</sub>, wherein R4 is as defined above.
- (E) R4-NH-(C=X)-R2, wherein all variables are as defined above.
- 15 (F) R4-NH-(C=X)-NH-Q, wherein Q is -R9-R10, wherein R9 is substituted or unsubstituted, alkyl, alkenyl, aryl, aralkyl, cycloalkyl and heterocycle, and R10 is a functional group selected from the group consisting of -NH<sub>2</sub>, -OH, -COOH, -COCl, -SH, -SO<sub>2</sub>Cl, -SO<sub>2</sub>H, -SO<sub>3</sub>-lower alkyl, provided that the terminal hydrogen, halogen or a reactive moiety of R10 may be replaced or protected by a
- 20 protecting group, -R8, and moiety Q is selected such that the terminal substituent thereof is reactive with the terminal substituent of T within reactant (G), permitting bond formation between (G) and (F), upon contact thereof.
- (G) T-Z-SS, wherein T is -CH<sub>2</sub>Cl, -NH<sub>2</sub>, or -COOH.
- (H) NH<sub>2</sub>-R7, wherein R7 is as defined above.
- 25 (J) R4-NH-(C=O)-NH-Q-R8, wherein all variables are as previously defined.
- (K) R4-NH-(C=O)-NH-Q, wherein all variables are as previously defined.
- (L) R4-NH-(C=S)-NH-Q-R8, wherein all variables are as previously defined.
- (M) R4-NH-(C-S-R11)-Q-R8, wherein all variables are as previously defined.
- (N) NH<sub>2</sub>-Q-R8, wherein Q and R8 are as previously defined.
- 30 (P) R1-(C=X)-NH-Q-R8, wherein all variables are as previously defined.

- (U)  $R4-NH-(C=X)-NH-Q-R8$ , wherein all variables are as previously defined.
- (W)  $HOOC-NH-Q-R8$ , wherein all variables are as previously defined.
- (AA)  $R4-N=C=X$ , an isocyanate when X is oxygen; a thiocyanate, when X is sulfur, and a carbodiimide, when X is N-R7.
- 5 (BB)  $(- )Y-Z-SS$ , an anion.

### Synthetic Route 1:

Synthetic Route 1 for production of the derivatized resin of formula (I):

- 10 (I)  $R4-NH-(C=X)-Y-Z-SS$   
comprises the steps of:
- (i) reacting a starting material of formula
    - (A)  $R-Y-Z-SS$   
with a reactant of formula
    - 15 (B)  $R1-(C=X)-R2$   
to form an intermediate product of formula
    - (C)  $R1-(C=X)-Y-Z-SS$ ;
  - (ii) reacting the intermediate (C) with a reactant of formula
    - (D)  $R4-NH_2$
    - 20 to form the product (I) of formula  $R4-NH-(C=X)-Y-Z-SS$ ; and
  - (iii) recovering the product (I).

### Synthetic Route 2:

- 25 Synthetic Route 2 for production of
- (I)  $R4-NH-(C=X)-Y-Z-SS$   
comprises the steps of:
- (i) reacting a starting material of formula
    - (B)  $R1-(C=X)-R2$
    - 30 with a reactant of

(D)  $R_4-NH_2$

to form an intermediate of formula

(E)  $R_4-NH-(C=X)-R_2$ ;

(ii) reacting intermediate (E) with a reactant of formula

5 (A)  $R-Y-Z-SS$

to form the product (I) of formula  $R_4-NH-(C=X)-Y-Z-SS$ ; and

(iii) recovering the product I.

### Synthetic Route 3:

10

Synthetic route 3 for production of

(I)  $R_4-NH-(C=X)-Y-Z-SS$

comprises the steps of:

(i) reacting a starting material of formula

15 (AA)  $R_4-N=C=X$ ,

with the an anion of formula (BB):

(BB)  $(-)Y-Z-SS$ ,

and quenching the reaction with a proton source to form the product (I).

20 For carrying out this embodiment of the invention, it will be appreciated that a wide variety of proton sources may be employed, including but not limited to strong or weak acids, such as water, HCl, acetic acid and the like. In reactant (AA), where X is oxygen, an isocyanate reactant is utilized. Where X is sulfur, an appropriate thiocyanate is utilized, and where X is N-R<sub>7</sub>, a commercially available carbodiimide is employed.

25

The anion Y(-) in reactant (BB) is generated to permit reaction with reagent (AA) according to methods known in the art for anion, including carbanion, generation, including but not limited to treatment with an appropriate base. For this purpose, the base may be selected from the group consisting of: sodium or potassium hydride, sodium

potassium or lithium alkoxide (e.g. ethoxide), potassium tert-butoxide, lithium diisopropyl amide, lithium or potassium bis(trimethylsilyl) amide, and the like.

#### Synthetic Route 4:

5

Synthetic route 4 for production of

(IA)  $R_4-NH-(C=X)-NH-Z-SS$

comprises the steps of:

(i) reacting a starting material of formula :

10

(G)  $T-Z-SS$

with a reactant of formula:

(F)  $R_4-NH-(C=X)-NH-Q$ ;

to form the product (IA);

(ii) recovering the derivatized resin (IA).

15

The reactant (F) of formula:

(F)  $R_4-NH-(C=X)-NH-Q$ ;

is prepared by a process comprising:

(i) reacting a starting material (N) of formula:

20

(N)  $H_2N-Q-R_8$

with a reactant (B) of formula:

(B)  $R_1-(C=X)-R_2$

to form an intermediate (P) of formula:

(P)  $R_1-(C=X)-NH-Q-R_8$ ;

25

(ii) reacting intermediate (P) with reactant (D) of formula

(D)  $R_4-NH_2$

to form an intermediate (U) of formula:

(U)  $R_4-NH-(C=X)-NH-Q-R_8$ ; and

(iii) removing  $R_8$  to form reactant (F);

30

wherein all variables are as previously defined.

- Depending on the nature of the functional group(s) comprising Q, those skilled in the art will appreciate that the nature of the protecting group -R8 may need to be varied, (see the discussion below which describes preferred combinations of protecting groups and
- 5 functional groups, as well as methods of deprotection). Accordingly, -R8 may be the same or different than -R3.

c. Embodiments and Variations on Synthetic Routes 1-4 for Production of the Derivatized Resin (I) of Formula R4-NH-(C=X)-Y-Z-SS:

10

Various embodiments of the invented synthetic routes described above include the following methods, the specifics of which are dictated by the desirable properties of the final product (I), including the desired character of the variables R4, X, Y and Z.

- 15 Thus, for example, an embodiment of Synthetic Route 1, wherein Y is selected to be -NH-, produces a product, referred to as (IA), by a process, process (IA), which comprises the steps of:

(i) reacting a starting material of formula:

(A)  $\text{NH}_2\text{-Z-SS}$

20

with a reactant of formula:

(B)  $\text{R1-(C=X)-R2}$ , wherein R1 and R2 are independently selected leaving groups that may be the same or different, to form the intermediate (C) of formula:

(C)  $\text{R1-(C=X)-NH-Z-SS}$  ;

25

(ii) reacting the intermediate (C) with a reactant of formula:

(D)  $\text{R4-NH}_2$

to form the product (IA); and

(iii) recovering the product (IA).

It will be appreciated that the following specific embodiments of Synthetic Routes 1-4 may be conducted as follows:

When Y is selected to be absent or  $-\text{CH}_2-$ , and X is O, a product referred to as (IB) is prepared by process (IB) comprising the steps of:

- (i) reacting a starting material of formula:
  - (C)  $\text{R1}-(\text{C}=\text{X})-\text{Y}-\text{Z}-\text{SS}$ , wherein R1 is  $-\text{OH}$ , and X is O, with a reactant of formula:
    - (D)  $\text{R4}-\text{NH}_2$
- to form the product (IB) of formula:
  - $\text{R4}-\text{NH}-(\text{C}=\text{O})-\text{Y}-\text{Z}-\text{SS}$ ; and
- (ii) recovering the product (IB).

When Y is selected to be absent or  $-\text{CH}_2-$ , and X is S, a product referred to as product (IC), is prepared by process (IC) which comprises the steps of:

- (i) reacting a starting material of formula:
  - (C)  $\text{R1}-(\text{C}=\text{X})-\text{Y}-\text{Z}-\text{SS}$ , wherein R1 is  $-\text{OH}$ , and X is O, with a reactant of formula:
    - (D)  $\text{R4}-\text{NH}_2$
- to form the product (IB) of formula:
  - $\text{R4}-\text{NH}-(\text{C}=\text{O})-\text{Y}-\text{Z}-\text{SS}$ ;
- (iii) thionating the product (IB) to form the product (IC) of formula:
  - $\text{R4}-\text{NH}-(\text{C}=\text{S})-\text{Y}-\text{Z}-\text{SS}$ ,

provided that if Y is absent, Z does not comprise an immediately adjacent carbonyl; and

- (iv) recovering the product (IC).

Those skilled in the art will appreciate that a wide variety of thionating conditions may be employed in the thionating step of this procedure. For example, Lawesson's reagent (also referred to as "LR", which is a thionation reagent well known in the art), or  $\text{P}_2\text{S}_5$  in mild

base (e.g. pyridine or sodium bicarbonate), or like means, may be employed to achieve this result.

When Y is selected to be absent or  $-\text{CH}_2-$ , and X is NR7, a product referred to as product

5 (ID), is prepared by process (ID) which comprises the steps of:

(i) reacting a starting material of formula:

(C)  $\text{R1}-(\text{C}=\text{X})-\text{Y}-\text{Z}-\text{SS}$ , wherein R1 is  $-\text{OH}$ , and X is O, and if Y is absent, Z does not comprise an immediately adjacent carbonyl, with a reactant of formula:

10 (D)  $\text{R4}-\text{NH}_2$

to form the product (IB) of formula:

$\text{R4}-\text{NH}-(\text{C}=\text{O})-\text{Y}-\text{Z}-\text{SS}$ ;

(iii) thionating the product (IB) to form the product (IC) of formula:

$\text{R4}-\text{NH}-(\text{C}=\text{S})-\text{Y}-\text{Z}-\text{SS}$ ;

15 (iv) reacting the product (IC) with an alkylating agent capable of contributing an alkyl group R11, which is preferably lower alkyl to form the intermediate (H) of formula:

(H)  $\text{R4}-\text{N}=(\text{C}-\text{S}-\text{R11})-\text{Y}-\text{Z}-\text{SS}$ ; and

(v) reacting the intermediate (H) with  $\text{NH}_2-\text{R7}$ , to form the product (ID) of  
20 formula:

$\text{R4}-\text{NH}-(\text{C}=\text{NR7})-\text{Y}-\text{Z}-\text{SS}$ ;

(v) recovering the product (ID).

It will be appreciated that a wide variety of alkylating conditions may be employed to  
25 achieve the desired alkylation. For example, the alkylating agent may be selected from the group consisting of iodomethane, iodoethane, methylbromide, ethylbromide, allylbromide, allylchloride, dimethylsulfate,  $\text{CH}_3\text{OSO}_2\text{CF}_3$ , and the like.

While it will be appreciated that reagents are available wherein each synthetic route may  
30 be conducted with X being oxygen, sulfur or NR7, wherein reagent (B), for example, is

- selected to be a thiodiimidazole or thiophosgene, or an imine diimidazole. Thus, according to Synthetic Route 2, reactant (E) is prepared by a process wherein X of reactant (B) is a sulfur or wherein X is NR7. When X is an oxygen, the oxygen in the derivatized resin (I)  $R4-NH-(C=X)-Y-Z-SS$  produced by any of the Synthetic Routes disclosed herein may be converted to a sulfur or -NR7 by thionation, to derive X as a sulfur and optionally alkylating the sulfur followed by contacting with  $NH_2-R7$ , to produce the product wherein X is NR7. From the foregoing disclosure, it will also be appreciated by those skilled in the art that the product wherein X is NR7 may also conveniently be derived by directly reacting a starting material wherein X comprises NR7, which comprises the steps of:
- (i) reacting a starting material of formula  
 $(C) \quad OH-(C=NR7)-Y-Z-SS,$   
with a reactant of formula:  
 $(D) \quad R4-NH_2$   
to form the product (ID) of formula:  
 $R4-NH-(C=NR7)-Y-Z-SS;$  and
  - (ii) recovering the product (ID).
- In carrying out this embodiment of the synthetic method, the starting material (C) wherein R1 is -OH, X is NR7, and Y is not absent, may be prepared, for example, by contacting a reactant of formula (B):
- (B)  $R1-(C=X)-R2$ , wherein X is NR7, and R1 is -OH,  
with a starting material of formula (G):  
 $(G) \quad T-Z-SS,$   
to form the starting material (C) wherein X is NR7, R1 is -OH, and the reaction of T with reactant (B) results in formation of the moiety Y.

For purposes of this aspect of the invention, the reagent (B) preferably is or is formed from a diimidazole imine, such as phosgeneimine diimidazole.



As will be appreciated from the foregoing disclosure, products (IE, IF, and IG) may also be prepared according to the following embodiments of Synthetic Route 4, comprising the steps of:

(i) reacting a starting material of formula (W):

5 (W)  $\text{HOOC-NH-Q-R8}$   
with reactant (D) of formula

(D)  $\text{R4-NH}_2$   
to form intermediate (J) of formula:

(J)  $\text{R4-NH-(C=O)-NH-Q-R8}$ ;

10 (ii) removing R8 from intermediate (J) to form intermediate (K) of formula:

(K)  $\text{R4-NH-(C=O)-NH-Q}$ ; and

(iii) reacting the intermediate (K) with material (G) of formula:

(G)  $\text{T-Z-SS}$  to form the product (IB) of formula:

15 (IE)  $\text{R4-NH-(C=O)-NH-Z-SS}$ , wherein all variables are as previously defined.

Where X is S, prior to removal of R8, intermediate (J) is thionated to form intermediate (L) of formula:

20 (L)  $\text{R4-NH-(C=S)-NH-Q-R8}$ , which is then deprotected and reacted with T-Z-SS to form product (IF).

Likewise, where X is desired to be NR7, prior to deprotection, intermediate (L) is contacted with an alkylating agent capable of contributing an alkyl group R11 to form the intermediate (M) of formula:

25 (M)  $\text{R4-N=(C-S-R11)-Q-R8}$ ; which is then reacted with (H)  $\text{NH}_2\text{-R7}$ , deprotected and reacted with T-Z-SS (G) to form product (IG).

It will be appreciated that a wide variety of alkylating agents may be used to achieve this purpose. Preferably, the alkylating agent is selected from the group consisting of

iodomethane, iodoethane, methylbromide, ethylbromide, allylbromide, allylchloride, dimethylsulfate, and  $\text{CH}_3\text{OSO}_2\text{CF}_3$ .

As will be appreciated from the foregoing disclosure, the product (I) may be prepared by a number of specific variations of the general methods described above. By way of summary, it will be appreciated that depending on the desired properties of product (I), different starting materials, reactants, and intermediates, as set forth above, may be reacted in various synthetic combinations. It will further be appreciated that in general, where the group "C=X" is "C=O", it is preferred for Y, Z or any other immediately adjacent group to not be a carbonyl group.

It will also be appreciated that when R4 is -OH, or -O-R3, (in which event R4 is referred to herein as R4'), the final product is useful in the production of hydroxamic ester compounds, which may be reduced to form an aldehyde, or to conduct other desirable chemistry, including but not limited to esterifying the resin at the free amino hydroxyl group.

d. Further Description of Key Reactants Utilized According to Synthetic Routes 1-4:

i. Reactants (A) and (G), Derivatives thereof, and Solid Supports Useful According to the Methods of this Invention:

Those skilled in the art will know, based on the extensive disclosure provided herein, which solid supports may be used to advantage according to the present disclosure. By way of specific guidance, however, it is noted that solid supports having a formula (A) R-Y-Z-SS, including, for example,  $\text{NH}_2$ -SS are commonly known, are commercially available and may be readily prepared by those of ordinary skill in the fields of combinatorial chemical synthesis and peptide synthesis. For instance, a variety of  $\text{NH}_2$ -SS resins are commercially available from Novabiochem, Advanced Chemtech, Bachem, and Peptide International, as well as other commercial sources. Specific resins that may

be derivatized according to the present invention include, but are not limited to water soluble polymers, cottons, sheets, glasses, fiberglasses, pins, polystyrene polymers and resins, each of which has at least one functional amine group. Especially preferred selections for  $\text{NH}_2\text{-SS}$  are resins, most especially aminomethylated polystyrene resin and  
5 4-methyl benzhydrylamine resin, which are available from commercial sources, including those listed above.

Solid supports of formula (G) T-Z-SS, as used according to specific synthetic methods disclosed herein, include the  $\text{NH}_2\text{-SS}$  support discussed above, as well as those having  
10 functional chloride or carboxylic acid groups in place of the functional amine group. Such supports are available from the commercial sources noted above, as well as other commercial sources, and may be readily prepared according to methods known in the art.

Polymeric resins having one or more of the following characteristics are suitable for use  
15 as solid supports according to the method of this invention:

1. Resins which are insoluble in polar aprotic solvents such as dimethyl formamide (DMF), N-methylpyrrolidone, tetrahydrofuran (THF), and other solvents conventionally used in solid-phase peptide synthesis (such as dichloromethane or methanol).
2. Resins which are microporous and which have high surface areas in polar aprotic  
20 solvents.
3. Resins which are capable of being functionalized with groups which can react with aldehyde or ketoamide groups to form a bond which is stable during the subsequent addition of amino acids (or amino acid analogs) to the N-terminal end of the growing peptide chain.
- 25 4. Resins which are stable in the presence of reagents such as TFA, diisopropylethyl amine, dicyclohexylcarbodiimide (DCC) and other reagents conventionally used in solid phase peptide synthesis.

ii. Reactant (B), of Formula R1-(C=X)-R2:

Reactant (B) is employed according to various embodiments of this invention, as described above. With respect to R1 and R2 of reactant (B), R2 is chosen for its ease of departure upon reaction with the solid support amino functional groups, and R1 is chosen for its ease of departure upon reaction with the amino group of reactant (D). R1 and R2 can be the same or different. Suitable R1 substituents include but are not limited to those in the group consisting of imidazole, p-nitrophenoxy, Cl, succinimidyl, and Me-imidazolium. Suitable R2 substituents include those in the group consisting of imidazole, Cl, succinimidyl, and Me-imidazolium. Preferred combinations of R1 and R2 include those where R1 and R2 are the same, or where R1 is p-nitrophenoxy and R2 is Cl. A preferred X in reactant B is N-alkyl, oxygen, sulfur, with oxygen being most preferred. When X is oxygen, an especially preferred reactant (B) is selected from the group consisting of 1,1-carbonyldiimidazole, p-nitrophenyl-chloroformate, phosgene, N,N'-disuccinimidyl carbonate, and 1,1-carbonyldimethyldiimidazolium. When X is NR7, a preferred reactant (B) is a diimidazole imine, such as phosgeneimine diimidazole. When X is sulfur, a preferred reactant (B) is 1,1'-thiocarbonyldiimidazole or thiophosgene. Reactant (B) of formula R1-(C=S)-R2 is chosen when functionalities in the final product other than carbonyl are desired. Accordingly, choice of this reactant permits the production of a product bearing the group -C=NR7, wherein R7 is a hydrogen or is an alkyl, alkenyl, aryl, aralkyl, cycloalkyl or heterocyclic substituent.

iii. Reactant (D) of Formula R4-NH2:

As defined above, R4 may be -OH, -NH2, -O-R3, or -NH-R3, wherein R3 is a protecting group. Suitable protecting groups are described below and are known in the art and can be found, for example, in "Protective Groups in Organic Synthesis", T. Greene, (John Wiley & Sons, Inc., 1991). When R4 is R3-NH-, preferred R3 protecting groups include Cbz, Boc, Alloc, Fmoc, Troc, Me3Si(CH2)2OCO, and PMC. Preferred species of reactant

(D) include  $\text{NH}_2\text{-NH}_2$  and  $\text{R}^3\text{-NH-NH}_2$ , wherein  $\text{R}^3$  is selected from the group consisting of Cbz, Boc, Alloc, Fmoc, Troc, and  $\text{Me}_3\text{Si}(\text{CH}_2)_2\text{OCO}$ .

iv. Protecting Groups:

5

It will be appreciated, particularly in the synthesis of peptides, peptide analogs and peptidomimetic compounds, that where reactive moieties are present, as in reactive amino acid side chains, if reactions at that reactive moiety is not desired, methods are known for protecting such sites. Accordingly, where a reactive heteroatom exists in an amino acid

10

side chain, or where reaction at an amino acid or amino acid analog's amino-terminal nitrogen is to be avoided, the following guidelines and procedures will apply. However, where side chains are non-reactive, those skilled in the art will appreciate that protection is not required (e.g. methylcysteine is already effectively protected; the phenylalanine side chain is relatively non-reactive). In particular, with regard to an argininal moiety,

15

wherein the side chain has a guanidino group (the arginine side chain is:  $\text{-(CH}_2\text{)}_3\text{-NH-(C=NH}_2^+\text{)-NH}_2$ , wherein the moiety  $\text{-NH-(C=NH}_2^+\text{)-NH}_2$  represents the guanidino functionality), single or double protection of the side chain may be desirable or essential in order to achieve a desired reaction at another side, e.g., at the amino-terminal nitrogen, without production of a plurality of unwanted side-chain reactions. It will also

20

be appreciated that while the argininal moiety exists in solution in various tautomeric forms, appropriate side-chain protection may be employed to keep the side chain in a linear form.

25

In practicing the methods of the present invention, the following considerations apply to the selection of  $\alpha$ -amino protecting groups, side chain protecting groups, and carboxy protecting groups, all of which are preferably used to protect sites on a peptide chain synthesized using the present derivatized resin. In selecting suitable  $\alpha$ -amino protecting groups (PG1) to be used during the synthesis of desirable peptidyl or peptidomimetic compounds using the resins of this invention, the  $\alpha$ -amino protecting group should:

- (i) render the  $\alpha$ -amino function inert under the conditions employed in the coupling reaction;
  - (ii) be readily removable after the coupling reaction under conditions that will not remove side chain or carboxy terminus protecting groups; and
- 5 (iii) eliminate the possibility of racemization upon activation prior to coupling.

A suitable  $\alpha$ -amino protecting group may be selected from the group consisting of acid labile  $\alpha$ -amino protecting groups (cleavage conditions are noted in brackets []):

- (a) Triphenylmethyl (trityl) group is cleaved under very mild acid conditions [1% TFA].
- 10 (b) tert-Butyloxy carbonyl (Boc), t-amyloxycarbonyl, adamantyloxycarbonyl, 4-methoxy benzyloxycarbonyl; these protecting groups require moderately strong acids such as trifluoroacetic acid, hydrochloric, or boron trifluoride in acetic acid for their removal.
- (c) Benzyloxycarbonyl (CBz), 2-chlorobenzyloxycarbonyl (2-ClZ),
- 15 cycloalkyloxycarbonyl, and isopropylloxycarbonyl, require stronger acids, such as hydrogen fluoride, hydrogen bromide or boron trifluoroacetate in trifluoro acetic acid for their removal. The CBz and the 2-ClZ groups may also be cleaved by hydrogenation under palladium on carbon in methanol. A suitable  $\alpha$ -amino protecting group also may be selected from the group consisting of base labile  $\alpha$ -amino protecting groups. For instance, Fluorenylmethyloxycarbonyl (Fmoc) may be cleaved
- 20 by using 20% piperidine/DMF or excess diethylamine in THF. Allyloxycarbonyl (Alloc) may be cleaved by Pd (0) catalyst transfer of the allyl group to a nucleophile such as morpholine, dimedone, tributyl tin hydride and N-methyl aniline. Preferred  $\alpha$ -amino protecting groups (PG) include Boc, Fmoc, Alloc, and Cbz.

- 25 An amino acid side-chain protecting group should:
- (i) render the protected side chain functional group inert under the conditions employed in the coupling reaction;
  - (ii) be stable under the conditions employed in removing the  $\alpha$ -amino or the carboxy terminus protecting groups, and

(iii) be readily removable upon completion of the desired peptide under reaction conditions that will not alter the structure of the peptide chain. A suitable amino acid side chain protecting group may be selected from the group consisting of (methods for cleavage of these protecting groups are shown in brackets [ ]):

- 5 (a) for protection of lysine amino groups, any of the groups mentioned above for the protection of  $\alpha$ -amino protecting groups are suitable.
- (b) for protection of arginine guanidino group, the preferred protecting groups include nitro [H<sub>2</sub>/Pd/C, HF], benzyloxycarbonyl (CBz) [HF, TFMSA, TMSOTf, H<sub>2</sub>/Pd/C], tert-butyloxycarbonyl (Boc) [TFA], 2,2,5,7,8-pentamethylchroman-6-sulfonyl (Pmc) [TFA], 2,3,6-trimethyl-4-methoxyphenylsulfonyl (Mtr) [TFA], p-toluenesulfonyl (Tos) [HF, TFMSA], mesitylene-2-sulphonyl (Mts) [HF, TFMSA], allyloxycarbonyl (Alloc) [Pd(0), morpholine or dimedone].
- 10 (c) For protection of serine and threonine hydroxyl groups, protecting groups include trityl [1% TFA], tert-butyl [TFA], benzyl, and substituted benzyl groups such as 4-methoxybenzyl, 4-chlorobenzyl, 2-chlorobenzyl, and 2,6-dichlorobenzyl which are cleaved by a similar method [HF, TFMSA, H<sub>2</sub>/Pd/C].
- 15 (d) For protection of tyrosine phenolic group, protecting groups such as tert-butyl [TFA], trityl [1% TFA], benzyl, 2-bromobenzyl and 2,6-dichlorobenzyl, all cleaved by the same reagents [HF, TFMSA, H<sub>2</sub>/Pd/C], are suitably employed.
- 20 (e) For protection of aspartic and glutamic acid side chain carboxy group, protecting groups include methyl [OH<sup>-</sup>, H<sup>+</sup>], ethyl [OH<sup>-</sup>, H<sup>+</sup>], t-butyl [TFA], allyl [Pd(0), morpholine], cyclohexyl [HF, TMSOTf], or benzyl groups [HF, TFMSA, TMSOTf, H<sub>2</sub>/Pd/C].
- (f) For protection of asparagine and glutamine side chain, protecting groups include trityl [TFA] and xanthyl [TFA].
- 25 (g) For protection of histidine imidazole group, suitable protecting groups include 2,4-dinitrophenyl (Dnp) [thiophenol], trityl [TFA], benzyloxymethyl (Bom) [HF, TFMSA, TMSOTf, H<sub>2</sub>/Pd/C], p-toluene sulfonyl (Tos) [HF, TFMSA], and benzyloxycarbonyl (Cbz) [HF, H<sub>2</sub>/Pd/C].

- (h) For protection of cysteine sulfhydryl group, suitable protecting groups include trityl [TFA], 4-methylbenzyl (pMeBzl) [HF, TFMSA], 4-methoxybenzyl (pMeOBzl) [HF, TFMSA], acetamidomethyl (Acm) [I2, Hg2+], tert-Butyl (tBu) [Hg2+].
- (i) For protection of tryptophan indole group, suitable protecting groups include formyl [10% piperidine in DMF, followed by HF] and tert-butyloxycarbonyl (Boc) [TFA].

A carboxy terminus protecting group (PG2) should:

- (i) render the protected functional group inert under the conditions employed in the coupling reaction,
- (ii) be stable under the conditions employed in removing the  $\alpha$ -amino or the side chain protecting groups, and
- (iii) be readily removable upon completion of the desired peptide under reaction conditions that will not alter the structure of the peptide chain. For the protection of the carboxy terminus of amino acids suitable protecting groups include methyl [OH-, H+], ethyl [OH-, H+], tert-butyl [TFA], benzyl [OH-, H2/Pd/C] and allyl [Pd(0), morpholine] groups.

e. Preferred Aspects of the Invented Methods for Production of the Resin (I):

- A preferred embodiment of the method of this invention comprises selection of aminomethylated polystyrene resin or 4-methyl benzhydrylamine resin as NH<sub>2</sub>-SS, reactant (A) wherein Z is absent, selection of reactant (B) from the group consisting of 1,1-carbonyldiimidazole, p-nitrophenylchloroformate, phosgene, N,N'-disuccinimidyl carbonate, and 1,1-carbonyldimethyldiimidazolium, and selection of -NH-R3 as R4 in reactant (D), with R3 selected from the group consisting of Cbz, Boc, Alloc, Fmoc, Troc, Me<sub>3</sub>Si(CH<sub>2</sub>)<sub>2</sub>OCO, and PMC. Aminomethylated polystyrene resin is most preferred.

- Where utilized, a resin of formula (G) T-Z-SS preferably comprises a functional amine group. Selection of T-Z-SS to be aminomethylated polystyrene resin or 4-methyl benzhydrylamine resin is especially preferred.

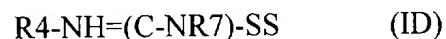
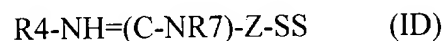
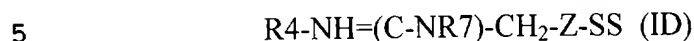
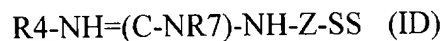


Where X in reactant (B) is sulfur, the selection of 1,1'-thiocarbonyl-diimidazole or thiophosgene as reactant (B), and the selection of NH<sub>2</sub> or R<sub>3</sub>-NH- as the R<sub>4</sub> group in reactant (D) is preferred for preparation of a solid support of formula R<sub>4</sub>-NH-(C=S)-NH-SS. When R<sub>4</sub> is selected to be R<sub>3</sub>-NH-, preferably, the R<sub>3</sub> protecting group is removed from the solid support thus formed to yield an -NH<sub>2</sub> terminal substituent on the solid support.

In a preferred embodiment, the product (I) is selected to have a formula NH<sub>2</sub>-NH-(C=O)-NH-SS, as disclosed in Examples 1, 2, and 3, and elsewhere throughout the specification. Reaction of an aldehyde or ketoamide with the -NH<sub>2</sub> moiety of the resin results in immobilization of the aldehyde or ketoamide. Subsequent chemical modification thereof, as in peptide synthesis, combinatorial chemistry and the like, results in production of a reaction product immobilized on the resin through a semicarbazone moiety. Release of the immobilized aldehyde or ketoamide, or a derivative thereof which is subjected to chemical modification while being immobilized on the resin, is thereafter achieved as disclosed herein, in view of which methods known in the art for cleavage of the semicarbazone moiety may be employed.

In view of the foregoing disclosure, it will be apparent that this invention contemplates methods of preparing a derivatized solid support having a formula including, but not limited to those represented by any of the following, with all variables as defined above:

- |    |  |      |
|----|--|------|
|    | R <sub>4</sub> -NH-(C=X)-Y-Z-SS                | (I)  |
|    | R <sub>4</sub> -NH-(C=X)-NH-Z-SS               | (IA) |
| 25 | R <sub>4</sub> -NH-(C=X)-NH-SS                 | (IA) |
|    | R <sub>4</sub> -NH-(C=O)-NH-Z-SS               | (IB) |
|    | R <sub>4</sub> -NH-(C=O)-CH <sub>2</sub> -Z-SS | (IB) |
|    | R <sub>4</sub> -NH-(C=O)-Z-SS                  | (IB) |
|    | R <sub>4</sub> -NH-(C=O)-SS                    | (IB) |
| 30 | R <sub>4</sub> -NH-(C=S)-NH-Z-SS               | (IC) |



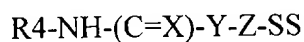
wherein SS is a solid support, and all other variables are as previously defined.

The invented solid supports prepared according to the synthetic methods disclosed herein  
 10 may be stored dry in a dessicator, preferably under refrigeration or at room temperature.  
 Resins prepared according to the invented methods are stable under these conditions for  
 six months or longer.

**B. Reacting an aldehyde or ketoamide with the derivatized resin of step (A),**

**15 thereby producing an immobilized aldehyde or ketoamide:**

Upon production, according to the above described method of this invention, of a suitably  
 derivatized resin of formula (I)



20 an appropriately protected aldehyde or ketoamide is reacted with the derivatized resin (I),  
 produced as described above in step (A). As such, the resin may be viewed as a  
 protecting group of the aldehyde or ketoamide moiety. The resin may also conveniently  
 be viewed as a tether or means for immobilization of the aldehyde or ketoamide.

Desirably, the aldehyde or ketoamide is reacted with the derivatized resin so as to  
 25 produce a product which may be later cleaved from the resin to produce a desired peptide  
 or peptidomimetic product, such as, for example, a pharmacologically significant protease  
 inhibitor being a terminal aldehyde or ketoamide moiety.

Upon production of the resin (I), wherein R4 is -NH-R3, the protecting group R3 is  
 30 removed to yield a free amino group. Contacting an appropriately protected aldehyde or

5 ketoamide with the free amino group, as described herein and as specifically exemplified hereinbelow, results in the formation of a semicarbazone moiety. In one preferred embodiment of the invention, the aldehyde is an appropriately protected argininal or an argininal peptide. Where an argininal is reacted with the resin, (i.e. the P1 position of a larger molecule is intended to be an argininal), and where further chemistry is to be conducted (e.g. peptide synthesis on the amino terminus of the argininal), it is preferred for the argininal to be orthogonally protected. In one preferred example of such orthogonal protection, it is preferred for the argininal side-chain amino groups to be di-Alloc or di-Boc protected, and the argininal amino terminal nitrogen to be Fmoc protected. In this manner, Fmoc peptide synthesis may be safely conducted, without the danger of deprotection of the argininal side-chain amino groups in the course of subsequent peptide synthesis steps at the argininal amino terminus. Furthermore, upon completion of the peptide synthetic steps, the peptide may conveniently be cleaved from the resin and the di-Boc or di-Alloc protection may be removed, in a single step, and under relatively mild acid-cleavage conditions, preferably including treatment with TFA; TFA:H<sub>2</sub>O, 9:1; TFA:DCM: H<sub>2</sub>O, 8:1:1; as opposed to HF treatment, relatively rapidly and with reduced danger of aldehyde dehydration to undesirable covalent adducts. Those skilled in the art will appreciate, based on this disclosure, that it is preferred for the lowest acid concentration and the shortest time to be used consistent with efficient cleavage and deprotection.

Those skilled in the art will appreciate that while the orthogonal protection described herein is preferred, under certain synthetic conditions, for example where harsh acidic conditions are required for synthetic steps subsequent to coupling of the aldehyde to the resin, the di-Boc or di-Alloc combined with Fmoc synthetic chemistry may result in premature deprotection of the argininal side chain amino groups. Accordingly, other known synthetic regimens would need to be employed under those circumstances.

It will also be appreciated from the foregoing disclosure that aldehyde or ketoamide isolation or purification may be achieved by contact with the derivatized resin produced

according to the method of this invention, and subsequent cleavage and recovery from the resin of the free aldehyde or ketoamide.

**C. Optionally performing solid-phase chemistry on the immobilized aldehyde or ketoamide of step (B), selected from peptide synthesis, synthesis of peptide analogs, and combinatorial chemistry:**

Once immobilized on the resin, solid-phase synthetic manipulation of aldehydes and ketoamides may be conducted. Standard solid-phase peptide synthetic methods may be employed, bearing in mind the specifics of the disclosure and preferences described herein. Thus, for example, it is preferred where a di-Boc argininal is desired to be directly coupled to the resin, Fmoc protection of the argininal amino-terminal nitrogen is employed. In this way, N-terminal chemistry may be conducted, without disrupting the side-chain protection.

According to the method disclosed herein, any of a wide variety of pharmacologically significant peptide or peptidomimetic compounds may be produced. Of particular significance is the ability to efficiently synthesize peptide aldehydes and ketoamides for use as protease inhibitors, including inhibitors of proteases significant to the blood coagulation pathway, including but not limited to thrombin inhibitors, factor Xa inhibitors and the like. Specific compositions prepared and disclosed according to this aspect of the present invention include, but are not limited to:

t-Butoxycarbonyl-Hydrazyl-Carbonyl-Amino Methylated polystyrene resin;

Hydrazyl-Carbonyl-Amino Methylated polystyrene resin;

$N^{\alpha}$ -Boc- $N^{\delta}$ -(di-Alloc)-Argininal-hydrazylcarbonyl-amino methylated polystyrene resin;

$N^{\delta}$ -(di-Alloc)-Argininal-hydrazylcarbonyl-amino methylated polystyrene resin;

Benzylsulfonamido-6Lactam-Gly- $N^{\delta}$ -(di-Alloc)-Argininal-hydrazylcarbonyl-amino methylated polystyrene resin;

Benzylsulfonamido-6Lactam-Gly-Argininal-hydrazylcarbonyl-amino methylated polystyrene resin;

N<sup>α</sup>-Boc-Gly(Allyl)al-HydrazylCarbonyl-Amino Methylated polystyrene resin;

N<sup>α</sup>- Gly(Allyl)al-HydrazylCarbonyl-Amino Methylated polystyrene resin;

5 hydrazonylcarbonyl aminomethylated polystyrene resin;

omega, omega'-di-N-t-butoxycarbonyl-Argininal hydrazonylcarbonyl aminomethylated polystyrene resin;

N-α-Fmoc-azetidine-2-carbonyl-omega, omega'-di-N-t-butoxycarbonyl-argininal hydrazonylcarbonyl aminomethylated polystyrene resin;

10 hydrazonylcarbonyl aminomethylated polystyrene resin;

N-α-Fmoc-seryl(O-t-Bu)-azetidine-2-carbonyl-N-omega, omega'-di-N-t-butoxycarbonyl-argininal hydrazonylcarbonyl aminomethylated polystyrene resin;

Seryl(O-t-Bu)-azetidine-2-carbonyl-omega, omega'-di-N-t-butoxycarbonyl-argininal hydrazonylcarbonyl aminomethylated polystyrene resin;

15 N-α-benzenesulfonyl-D-seryl(O-t-Bu)-azetidine-2-carbonyl-omega, omega'-di-N-t-butoxycarbonyl-argininal hydrazonylcarbonyl aminomethylated polystyrene resin;

Nα-Fmoc-N<sup>d</sup>-(di-Alloc)-Arginal-hydrazylcarbonyl-amino methylated polystyrene resin;

Those skilled in the art will appreciate from the foregoing disclosure that the derivatized

20 resins according to the present invention may be utilized in any of a number of useful ways. Included among the methods of using the present resins are methods for solid-phase synthesis of peptides, methods for conducting combinatorial chemistry and methods for purifying or isolating proteins or peptides. Those skilled in the art will appreciate that other methods for using the derivatized resins according to the instant

25 invention exist, and those methods likewise come within the scope of the present invention. Thus, for example, the resins according to the present invention are particularly useful in the solid-phase synthesis of protease inhibitors, and in particular inhibitors of serine proteases, such as thrombin and Factor Xa, urokinase, the NS3 protease of Hepatitis C virus and trypsin. Further specifics of these aspects of the method

of this invention for utilization of the derivatized resin (I) are provided in further detail below:

i. Solid-phase Peptide Synthesis:

5

One aspect of the present invention includes the use of the synthetic method of this invention, including the generic and specific synthetic methods disclosed herein, and compositions made thereby, (collectively referred to as "the invented solid supports"), in the solid phase synthesis of peptide or peptidomimetic compositions. Preferred uses include use for the synthesis of peptide or peptidyl aldehydes or ketoamides, such as those that inhibit one or more serine proteases. An especially preferred use of the invented solid supports is use for the synthesis of inhibitors of thrombin, Factor Xa, urokinase, Hepatitis C enzymes, chymase, prostate specific antigen (PSA), Factor VIIa, elastase, and trypsin.

10

15

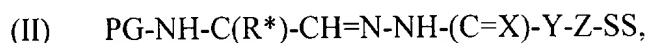
Examples 4-8 describe the use of synthetic methods disclosed herein to make a thrombin inhibitor. Examples 9-14 and 23-26 describe the use of the synthetic methods disclosed herein to make specific inhibitors of urokinase .

20

In a particularly preferred embodiment of this aspect of the invention, Boc protection of the aldehyde permits simultaneous deprotection of the peptide and cleavage of the peptide from the resin, to release the free aldehyde product.

25

According to this method of producing derivatized resins and use thereof to produce peptides of pharmacologic significance, R4 of the product (I), wherein R4 is R3-NH- is further derivatized such that a product is produced that may be represented by the formula (II):



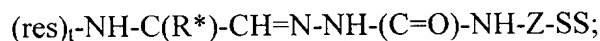
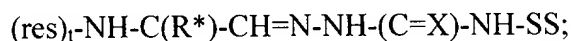
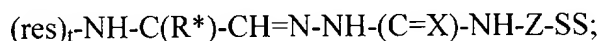
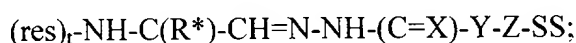
wherein (R\*) represents an amino acid side chain. Where an oligomer is produced according to this method, the product (I) is further derivatized at the R4 group to produce a product which may be represented by the formula (III):



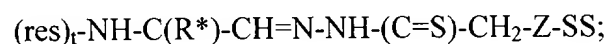
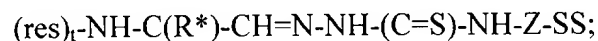
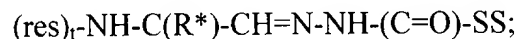
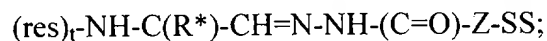
- 5 wherein (res)<sub>t</sub> represents a peptide chain of “t” residues, wherein “t” is an integer between about 2 and about 50, and more preferably between about 3 and 10.

As a result of practicing the method of this invention, therefore, it will be appreciated that a product, including but not limited to the following variations on the derivatized resin

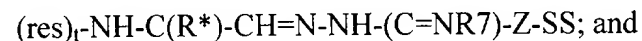
- 10 represented by formula (III), is produced:



- 15  $(res)_t-NH-C(R^*)-CH=N-NH-(C=O)-CH_2-Z-SS;$



- 20  $(res)_t-NH-C(R^*)-CH=N-NH-(C=S)-Z-SS;$



- 25  $(res)_t-NH-C(R^*)-CH=N-NH-(C=NR_7)-SS.$

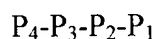
It will further be appreciated from the foregoing disclosure that the group (res)<sub>t</sub>, may be produced by standard solid-phase synthetic procedures known in the art wherein individual amino acid or amino acid analog residues are linked, one by one, to form the peptide or peptide analog. At each step, standard protection, deprotection, coupling and

wash steps are conducted. In an alternate embodiment of this aspect of the invented method, blocks of separately synthesized residues, i.e. oligomers or polymers, may be contacted with the product represented by formula (II), to produce a product of formula (III), in a reduced number of steps. Equivalents and other variations suggested to those skilled in the art based on the foregoing disclosure may likewise be conducted without departing from the invention disclosed and claimed herein.

Cleavage of the peptide from the resin and deprotection results in production of the desired peptide or peptidomimetic product produced by extension of the peptide chain on the derivatized solid support according to the method of this invention.

## ii. Combinatorial Chemistry:

The derivatized resin (I) produced according to the method of this invention is also useful in the field of combinatorial chemistry and development of chemical libraries. With reference to the following disclosure, it will be appreciated that preferred methods within this aspect of the invention result in the development of combinatorial libraries having an aldehyde or ketoamide in the P<sub>1</sub> position, especially argininals and glycinals:



Those skilled in the art will recognize that while a molecule bearing four residues is represented, peptides or peptide analogs of any desired length may be prepared according to this method by repeating the coupling steps as many times as necessary. According to one embodiment of this aspect of the invention, a library is designed wherein the P<sub>1</sub> residue may be varied or kept constant while residues P<sub>4</sub>, P<sub>3</sub>, and P<sub>2</sub> are varied and incorporated into, for example, peptides and peptidomimetic P<sub>1</sub> aldehydes. Using the invented supports, multiple reactions are carried out in parallel. The P<sub>1</sub> site is first incorporated onto the resin, e.g. the HCAM resin of this invention, following procedures exemplified in Examples 1-5, followed by incorporation of variations in the P<sub>4</sub>, P<sub>3</sub>, and P<sub>2</sub>



residues, thereby forming a library of peptide or peptidomimetic compounds available for structure-activity analyses in any of a number of *in vitro* or *in vivo* assay systems, including protease inhibition assays.

- 5 As will be appreciated by those skilled in the art, this process of library formation and parallel synthesis may be carried out in a number of known formats. In one embodiment, the synthesis is conducted in Whatman mini columns, or the like, wherein standard peptide synthetic methods known in the art are used to extend the peptide chain, with each subsequent coupling being achieved at the carboxy terminus of each added residue
- 10 which is linked to the amino terminus of the previously immobilized residues.

Following synthesis, the multiple peptide variants are cleaved from the resin, isolated, and tested for biological activity. Likewise, with the aid of the invented solid supports, automated synthesis of a library of peptides or peptide analogs may be conducted in

15 commercially available peptide synthesizers.

Thus, this aspect of the invention represents a method for making a library of peptides or peptide analogs comprising the steps of:

- (a) in each of a series of separate containers or reaction vessels, contacting an aldehyde or ketoamide with a derivatized resin represented by formula (I) under conditions which
- 20 permit the formation of a stable semicarbazone, thereby immobilizing said aldehyde or ketoamide to produce a P<sub>1</sub> residue;
- (b) in each of said series of separate containers or reaction vessels, contacting same or different amino acid residues or amino acid analog residues with the thus
- 25 immobilized aldehyde or ketoamide under conditions permitting formation of a peptide bond so as to produce a series of P<sub>2</sub> residues, same or different, in each of said series of separate containers or reaction vessels; and
- (c) repeating step (b) as many times as required so as to produce a sequential series of P residues to generate a peptide or peptide analog of the desired number of residues,

with appropriate intermediate steps of protection and deprotection of reactive groups present on the growing peptide or peptide analog chain.

Standard peptide synthetic methods may be employed in this process, subsequent to the initial formation of the aldehyde-resin or ketoamide-resin linkage, as described above. Subsequent to cleavage from the resin, the thus synthesized library of peptides or peptide analogs bearing the original aldehyde or ketoamide as the P<sub>1</sub> residue may then be purified and tested for biological activity.

iii. Peptide or Protein Purification and Separation:

A further use of the invented solid supports, which comprise an amine, is in the field of purification and separation. The solid supports can be used to bind to, and thereby immobilize and isolate from solution, any composition bearing a reactive carbonyl component. The solid support-bound, carbonyl containing component may be a contaminant, as thus removed to increase purity of the starting solution, or it may be a desired product, in which case binding serves as step in the purification of the product.

For instance, the invention contemplates a method of isolating an aldehyde- or ketoamide-containing compound or composition from a solution, comprising the steps of:

- (a) contacting a solution comprising an aldehyde or ketoamide with derivatized resin represented by formula (I) under conditions permitting interaction of the aldehyde carbonyl with the amine of the derivatized resin; and
- (b) allowing sufficient time for the interaction to proceed to completion such that the aldehyde or ketoamide is bound to the derivatized resin, thereby removing the aldehyde- or ketoamide-containing compound from the solution.

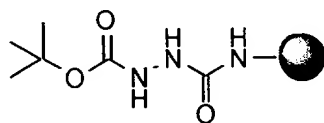
Optionally, the aldehyde or ketoamide can be removed from the derivatized resin by hydrolysis using procedures like those set forth in the specific exemplary support presented herein, and recovered by filtration in Whatman mini-columns or the like.

**D. Cleaving, deprotecting and recovering the product of step (C) as the free aldehyde or ketoamide.**

Based on the foregoing disclosure, those skilled in the art will appreciate that production of pharmacologically significant peptide or peptide analog products bearing an aldehyde or ketoamide in the P1 position is enabled by this disclosure. Upon completion of the peptide synthetic steps, the peptide may conveniently be cleaved from the resin. Where di-Boc or di-Alloc protection has been employed, as described above, the protecting groups may be removed and the peptide cleaved from the resin, in a single step, and under relatively mild acid-cleavage conditions (e.g. TFA; TFA:H<sub>2</sub>O, 9:1; TFA:DCM: H<sub>2</sub>O, 8:1:1; as opposed to HF treatment), relatively rapidly and with reduced danger of aldehyde dehydration to undesirable covalent adducts. Those skilled in the art will appreciate, based on this disclosure, that it is preferred for the lowest acid concentration and the shortest time to be used, consistent with efficient cleavage and deprotection. Depending on the type of protection, higher acid concentrations and cleavage/deprotection times may be required. Bearing this disclosure and specific guidance in mind, those skilled in the art are well aware of methods for peptide cleavage, deprotection and product recovery, which in this case permits facile recovery of product bearing a P1 aldehyde or ketoamide moiety.

**SPECIFIC EXEMPLARY SUPPORT:**

Having generally described the invention with respect to the preferred embodiments thereof, the following specific exemplary disclosure is provided. The specifics of these examples are provided to supplement the written description of this invention. However, the specifics of these examples should not be construed as limiting on the invention.

Example 1Preparation of t-Butoxycarbonyl-Hydrazyl-Carbonyl-Amino Methylated polystyrene resin

5

To a suspension of 1,1-carbonyldiimidazole (29.19 g, 180 mmol, 6 equiv.) in dimethylformamide (300 mL) was added portionwise t-butylcarbazate (23.76 g, 180 mmol, 6 equiv.) with stirring at ambient temperature under a nitrogen atmosphere. After the addition was completed, the reaction was allowed to stir at room temperature for 2 1/2

10

hours and then poured into amino methylated polystyrene resin (30 g, 30 mmol, 1 meq/g, Advanced Chemtech) in the peptide synthesis flask. The suspension was purged with nitrogen gas for 3 hours. The resin was filtered and washed with 3 X 350 mL portions of methylene chloride. Kaiser Test (solution and bead; light blue, clear). A double coupling was undertaken to ensure a quantitative reaction, although double coupling is optional.

15

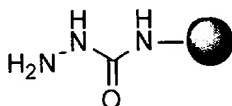
Thus, a solution of 1,1-carbonyldiimidazole (14.60 g, 90 mmol, 3 equiv.) and t-butylcarbazate (11.88 g, 90 mmol, 3 equiv.) in 150 dimethylformamide was stirred at ambient temperature for 30 minutes and then poured into the previously prepared resin. After purging with nitrogen gas for 1 hour, the resin was filtered and washed with 6 X 350 mL portions each of methylene chloride, methanol, methylene chloride, methanol, methylene chloride, and methanol. Kaiser Test (solution and bead; clear and clear).

20

Subsequent coupling reactions were carried out when necessary. The resin was dried under vacuum and acetylated with DMP/acetic anhydride/Et<sub>3</sub>N (8:1:1; about 300 mL) for 30 minutes at ambient temperature. The resin was successively washed three times each with solvents (methylene chloride and methanol) and dried under vacuum to afford

25

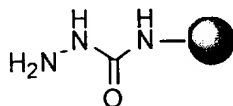
product in quantitative yield.

Example 2Hydrazyl-Carbonyl-Amino Methylated polystyrene**HCAM resin**

5

A solution of DCM/TFA (1:1) (300 mL) and thioanisole (10 mL) was added to the resin of Example 1 (30g) to effect deprotection. The mixture purged slowly with nitrogen gas in the peptide synthesizer flask at ambient temperature for 30 minutes. The deprotected resin was filtered and washed with DCM (2 X), DCM/DIEA (2 X), DCM (2 X), and alternatively with organic solvents (methylene chloride, methanol). The resulting title resin was dried under vacuum. Kaiser Test (solution and bead; light blue, sand color); Overall yield: 30.64 g resin (approx. 0.85 mmol/g or 0.85 meq semicarbazide/g).

10

15 Example 3Preparation of Hydrazyl-Carbonyl-Amino Methylated polystyrene resin**HCAM resin**

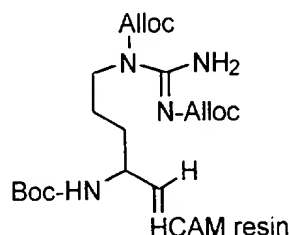
To amino methylated polystyrene resin (1.0 g, 1.0 mmol, Advanced Chemtech) in N,N-dimethylformamide (12 mL) was added 1,1-carbonyldiimidazole (3.89 g, 24.0 mmol, 24 equiv.). The suspension was shaken at room temperature for 5 hours and then filtered in a peptide synthesis flask. The resin was washed with copious amounts of methylene chloride. Kaiser Test (solution and bead; light blue, white). A 2M solution of hydrazine in N,N-dimethylformamide (12 mL) was added to the resin and allowed to shake at ambient temperature overnight. The resin was filtered and washed alternately with

20  
25

organic solvents (methylene chloride and methanol) and dried under vacuum to afford product 1.06 g (approx. 0.94 mmol/g).

#### Example 4

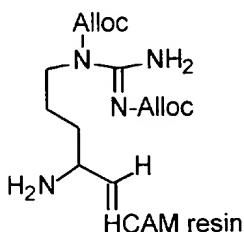
- 5 Preparation of N<sup>α</sup>-Boc-N<sup>δ</sup>-(di-Alloc)-Argininal-hydrazylcarbonyl-amino methylated polystyrene resin.



- 10 t-butoxycarbonyl-L-Arg(alloc)<sub>2</sub>-al (129 mg, 0.30 mmol, 2 equiv.; see Examples 15 and 16 for synthesis) was added to a suspension HCAM resin of Example 2 or 3 (200 mg, 0.15 mmol) in methylene chloride (2.52 mL) in a Whatman mini-column tube. The mixture was allowed to shake for 24 hours at ambient temperature. The resin was filtered and washed with copious amounts of solvents (methylene chloride, methanol, and ether).
- 15 Kaiser Test (solution and bead; clear and clear). The resin was dried under vacuum to afford 213 mg resin (approx. 0.57 mmol/g).

#### Example 5

- 20 Preparation of N<sup>δ</sup>-(di-Alloc)-Argininal-hydrazylcarbonyl-amino methylated polystyrene resin

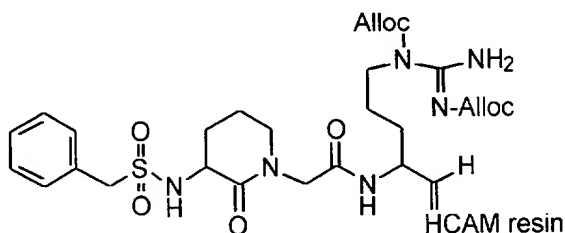


A solution of DCM/TFA/Thioanisole (7: 3: 1) (5 mL) was added to the resin of Example 4 (210 mg, 0.12 mmol). The mixture was allowed to shake in a Whatman mini-column tube at ambient temperature for 15 minutes. The deprotected resin was filtered and washed with DCM (3 X), DCM/DIEA (2 X), DCM (2 X), MeOH (2 X), DCM (3 X). The resulting title resin was dried under vacuum. Kaiser Test (solution and bead; blue and light brown); Resin substitution (0.61 mmol/g).

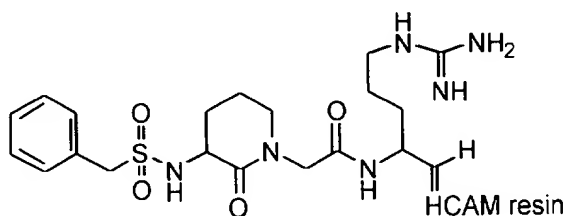
### Example 6

#### Preparation of Benzylsulfonamido-6Lactam-Gly-N<sup>δ</sup>-(di-Alloc)-Argininal-

hydrazylcarbonyl-amino methylated polystyrene resin



To a suspension of the resin of Example 5 (70 mg, 0.04 mmol), N-benzylsulfonyl-norVal(cyclo)-Gly-OH (19.6 mg, 0.06 mmol, 1.4 equiv.; see Example 11 of US 5,703,208 for synthesis) and PyBOP (31.3 mg, 0.06 mmol, 1.4 equiv.) in DMF (0.8 mL), was added DIEA (7.8 mg, 0.06 mmol, 1.4 equiv.). The mixture was allowed to shake in a Whatman mini-column at ambient temperature overnight. The resin was filtered and washed with organic solvents (methylene chloride and methanol) and dried. Kaiser Test (solution and bead; clear and clear). Based on theoretical yield (83 mg), Resin substitution (0.51 mmol/g).

Example 7Preparation of Benzylsulfonamido-6Lactam-Gly-Argininal-hydrazylcarbonyl-amino methylated polystyrene resin

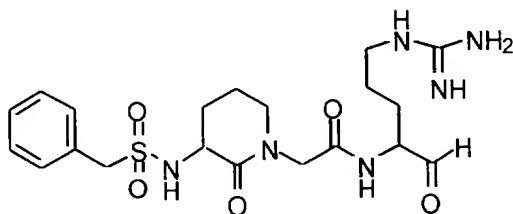
5

To a solution of THF/DMSO/.5N HCl/Morpholine (2: 2: 1: 5) (1.5 mL) was added the resin of Example 6 (83 mg, 0.043 mmol) and (Ph<sub>3</sub>P)<sub>4</sub> Pd (33 mg, 0.65 equiv.). The mixture was allowed to shake in a Whatman mini-column at room temperature for 4 hours. The resin was filtered and washed with organic solvents (methylene chloride and methanol) and dried. Based on theoretical yield (76 mg), Resin substitution was estimated to be (0.56 mmol/g).

10

Example 8Preparation of Benzylsulfonamido-6Lactam-Gly-Argininal

15



A solution of TFA/H<sub>2</sub>O (8: 2) (1.5 mL) was added to the resin of Example 7 (75 mg, 0.039 mmol) in a Whatman mini-column. The mixture was allowed to shake by mechanical shaker for 1 1/3 hours at ambient temperature. The filtrate was purified with a Sepak column, using a H<sub>2</sub>O, 0.1% acetonitrile; 10, 0 to 5, 5 gradient to afford 9.1 mg (50.0% yield) of title product as a white-powder (fluffy).

20

MS (M + H) = 467.0, calculated (MW) = 465.6



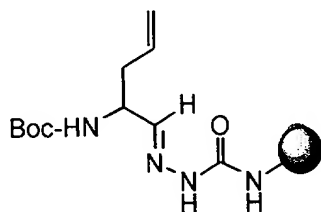
HPLC retention, 10.9, 11.8, and 13.0 min. (2 cyclol and hydrate forms of the argininal derivative); C18 5 - 50% MeCN over 25 minutes, 1.0 mL/min.

The title compound was shown to have thrombin inhibitory activity in *in vitro* assays.

5

#### Example 9

#### Preparation of N<sup>α</sup>-Boc-Gly(Allyl)al-Hydrazyl Carbonyl-Amino Methylated polystyrene resin



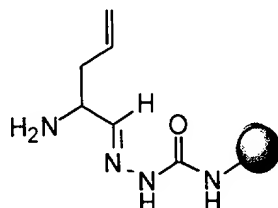
10

N<sup>α</sup>-Boc-Gly(Allyl)al (237 mg, 1.19 mmol, 1.4 equiv.; see Examples 17 through 19 for synthesis) was added to a suspension of HCAM resin of Example 2 or 3 (1 g, 0.85 mmol) in methylene chloride (10 mL). The mixture was allowed to stir slowly for 24 hours at ambient temperature. The resin was filtered and washed with copious amounts of solvents (methylene chloride, methanol). The resin was dried under vacuum (a 2 mg portion of the resin was taken for a Kaiser Test), and then acetylated with DMF/acetic anhydride/Et<sub>3</sub>N (8: 1: 1) for 20 minute at room temperature in order to ensure no free amine remained on the beads. Once again, the resin was dried under vacuum. Based on theoretical yield (1.15 g), Resin substitution (0.73 mmol).

15

20

25

Example 10Preparation of N<sup>α</sup>- Gly(Allyl)al-HydrazylCarbonyl-Amino Methylated polystyrene resin

5

A solution of DCM/TFA/Thioanisole (6: 4: 1) (15 mL) was added to the resin of Example 9 (1.15 g, 0.73 mmol). The mixture was allowed to shake in a Whatman mini-column tube at ambient temperature for 25 minutes. The deprotected resin was filtered and washed with DCM (3 X), DCM/DIEA (3 X), and alternately with organic solvents (methylene chloride and methanol). The resulting title resin was dried under vacuum to afford 1 g resin. Kaiser Test (solution and bead; dark purple and dark brown); Resin substitution (0.79 mmol/g).

10

The title resin was used as starting material to synthesize a serine protease inhibitor having a glycine aldehyde as its terminal residue.

15

Example 11Preparation of N-alpha-fluorenyloxycarbonyl-omega, omega'-di-N-t-butoxycarbonyl-arginine methyl ester:

20

To a solution of N-alpha-fluorenyloxycarbonyl-omega, omega'-di-N-t-butoxycarbonyl-arginine (4.97g, 8.329mmol, Advanced Chemtech) in acetonitrile (82mL) was added potassium carbonate (1.38g, 10mmol) and methyl iodide (1.037mL, 16.66mmol). The mixture was warmed to 50°C. After stirring for 4.5 hours at 50°C, the reaction mixture was poured into ethyl acetate (500mL) and washed successively with water (1x50mL), saturated sodium bicarbonate (1x50mL), and brine (1x50mL). The organic phase was

25

dried with sodium sulfate, and solvent was removed under reduced pressure yielding 4.7g of the title compound.

### Example 12

#### 5 Preparation of N-alpha-fluorenyloxycarbonyl-omega, omega'-di-N-t-butoxycarbonyl-argininol:

To a solution of the compound of Example 11 (5.675g, 9.29mmol) in tetrahydrofuran (15.5mL) and methanol (93mL) was added calcium chloride(2.06g, 18.58mmol), and the  
 10 mixture was cooled in an ice bath. Sodium borohydride (1.4g, 37.17mmol) was added slowly in portions to the stirred cooled solution. After 1.5h solvent was removed under reduced pressure. The residue was suspended in ethyl acetate (500mL) and water (300mL). The phases were separated, and the organic layer was washed successively with 0.5M HCl (200mL), and brine (200mL). The combined aqueous washes were back  
 15 extracted with ethyl acetate. The ethyl acetate layers were combined, silica was added (30g), and solvent was removed under reduced pressure. This silica was loaded onto a 150x80mm silica flash column, and the product was eluted with 50% ethyl acetate/hexanes to afford 3.855g (71% yield) of the title compound. 0.938g (17%) of the compound of Example 11 (starting material) was recovered. Rf=0.30 (50% ethyl  
 20 acetate/hexanes). <sup>1</sup>H NMR (CDCl<sub>3</sub>): 8.35ppm(bs, 1H), 7.75ppm(m, 2H), 7.6ppm(m, 2H), 7.38ppm(m, 2H), 7.3ppm(m, 2H), 5.45ppm(m, 1H), 4.4ppm(d, 2H), 4.2ppm(m, 1H), 3.7ppm(m, 2H), 3.57ppm(m, 1H), 3.45ppm(m, 1H), 3.35ppm(m, 2H), 2.15ppm(m, 1H), 1.6ppm(m, 4H), 1.45ppm(d, 18H).

### 25 Example 13

#### Preparation of N-alpha-fluorenyloxycarbonyl-omega, omega'-di-N-t-butoxycarbonyl argininal hydrazonylcarbonyl aminomethylated polystyrene resin:

To a solution of the compound of Example 12 (3.62g, 6.21mmol) in methyl sulfoxide  
 30 (31mL) and toluene (31mL) cooled in an ice bath was added EDC (11.93g, 62.1mmol)

and dichloroacetic acid (2.56mL, 31.06mmol). The reaction was stirred for 40 minutes until no starting material was observed by TLC (50% ethyl acetate/hexanes). Water (150mL) and ethyl acetate (500mL) were added, and the phases were separated. The organic layer was then washed successively with 0.5M HCl (100mL), saturated sodium bicarbonate (2x100mL), and brine (100mL). The organic layer was dried with sodium sulfate, and solvent removed under reduced pressure. This residue was then dissolved in dichloromethane (45mL), added to the solid support of Example 2 (3.65g, 3.1mmol), and stirred in a sealed tube end-over-end overnight. The resin was filtered and washed successively with dichloromethane (3x50mL), dimethylformamide (3x50mL), and methanol (3x50mL), and dried under high vacuum. Yield of title compound was 4.3g. The piperidine fulvene determination of the loading showed 0.37mmol/g (60% coupling).

#### Example 14

##### Preparation of benzenesulfonyl-D-seryl-azetidyl-argininal, trifluoroacetate salt

15 Step 1: omega, omega'-di-N-t-butoxycarbonyl-Argininal hydrazonylcarbonyl aminomethylated polystyrene resin

The resin product of Example 13, (125 mg) was treated with 30% piperidine/dimethylformamide (1.5mL) in a 4mL polypropylene column. After 30 min, the resin was drained, washed successively with dichloromethane (2X3mL) and methanol (2x3mL), and dried under vacuum to give the title compound. A ninhydrin assay on a small aliquot gave dark blue resin and solution showing a high yield for the deprotection.

25 Step 2: Preparation of N- $\alpha$ -Fmoc-azetidine-2-carbonyl-omega, omega'-di-N-t-butoxycarbonyl-argininal hydrazonylcarbonyl aminomethylated polystyrene resin:

To the compound of Step 1 (0.12g) in a 4mL polypropylene column were added N- $\alpha$ -Fmoc-azetidine-2-carboxylic acid (110mg, 0.34mmol), 1-hydroxybenzotriazole (7.7mg, .057mmol), TBTU (37mg, 0.11mmol) and diisopropylethylamine (29.8 $\mu$ L, 0.17mmol) in dimethylformamide (95 $\mu$ L). The reaction mixture was shaken at room temperature

overnight. The reagents were drained from the resin and the resin was washed successively with dichloromethane (2X3mL), and methanol (2X3mL). The resin was vacuum dried and a small aliquot was taken for ninhydrin colorimetric analysis which showed a 97% coupling efficiency in the production the title compound.

5

Step 3: Azetidine-2-carbonyl-omega, omega'-di-N-t-butoxycarbonyl-Argininal hydrazonylcarbonyl aminomethylated polystyrene resin

10 The compound of Step 2 was treated with 30% piperidine/dimethylformamide (1.5mL) in a 4mL polypropylene column. After 30 min, the resin was drained, washed successively with dichloromethane (2X3mL) and methanol (2x3mL), and dried under vacuum to give the title compound. A ninhydrin assay on a small aliquot gave dark blue resin and solution showing a high yield for the deprotection.

15 Step 4: Preparation of N- $\alpha$ -Fmoc-seryl(O-t-Bu)-azetidine-2-carbonyl-N-omega, omega'-di-N-t-butoxycarbonyl-argininal hydrazonylcarbonyl aminomethylated polystyrene resin:

To the compound of Step 3 (0.12g) in a 4mL polypropylene column were added N- $\alpha$ -Fmoc-D-serine t-butyl ether (43mg, 0.11mmol), 1-hydroxybenzotriazole (7.7mg, .057mmol), TBTU (37mg, 0.11mmol) and diisopropylethylamine (29.8 $\mu$ L, 0.17mmol) in dimethylformamide (95 $\mu$ L). The reaction mixture was shaken 4h at room temperature. The reagents were drained from the resin, and the resin was washed successively with dichloromethane (2X3mL), and methanol (2X3mL). The resin was dried under vacuum, then double coupled using N- $\alpha$ -Fmoc-serine O-t-butyl ether (43mg, 0.11mmol), 1-  
25 hydroxybenzotriazole (7.7mg, .057mmol), TBTU (37mg, 0.11mmol) and diisopropylethylamine (29.8 $\mu$ L, 0.17mmol) in dimethylformamide (95 $\mu$ L). The reaction mixture was shaken 2h at room temperature. The reagents were drained from the resin, and the resin was washed successively with dichloromethane (2X3mL), and methanol (2X3mL). The resin was dried under vacuum, and a small aliquot was taken for ninhydrin

colorimetric analysis which showed a 97.4% coupling efficiency in the production the title compound.

5 Step 5: Seryl(O-t-Bu)-azetidine-2-carbonyl-omega, omega'-di-N-t-butoxycarbonyl-argininal hydrazonylcarbonyl aminomethylated polystyrene resin

10 The compound of Step 4 was treated with 30% piperidine/dimethylformamide (1.5mL) in a 4mL polypropylene column. After 30 min, the resin was drained, washed successively with dichloromethane (2X3mL) and methanol (2x3mL), and dried under vacuum to give the title compound. A ninhydrin assay on a small aliquot gave dark blue resin and solution showing a high yield for the deprotection.

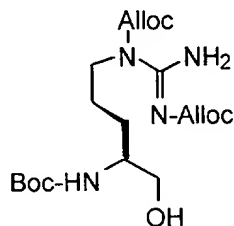
15 Step 6: Preparation of N- $\alpha$ -benzenesulfonyl-D-seryl(O-t-Bu)-azetidine-2-carbonyl-omega, omega'-di-N-t-butoxycarbonyl-argininal hydrazonylcarbonyl aminomethylated polystyrene resin:

20 To the compound of Step 5 (0.12g) in a 4mL polypropylene column were added benzenesulfonyl chloride (17 $\mu$ L, 0.133mmol), and diisopropylethylamine (46.3 $\mu$ L, 0.27mmol) in dimethylformamide (888 $\mu$ L). The reaction mixture was shaken overnight at room temperature. The reagents were drained from the resin, and the resin was washed successively with dichloromethane (2X3mL), and methanol (2X3mL). The resin was dried under vacuum, then double coupled with benzenesulfonyl chloride (17 $\mu$ L, 0.133mmol), and diisopropylethylamine (46.3 $\mu$ L, 0.27mmol) in dimethylformamide (888 $\mu$ L). The reaction mixture was shaken 3h at room temperature. The reagents were  
25 drained from the resin, and the resin was washed successively with dichloromethane (2X3mL), and methanol (2X3mL). The resin was dried under vacuum, and a small aliquot was taken for ninhydrin colorimetric analysis which showed a 96% coupling efficiency in the production the title compound.

30

Step 7: benzenesulfonyl-D-seryl-azetidine-2-carboxyl-argininal, trifluoroacetate salt

The compound of Step 6 (45mg) in a 4mL polypropylene fritted column was treated with trifluoroacetic acid/water (0.5mL of a 9:1 mixture). The column was shaken for 1.5 h at room temperature. The reaction solution was drained into a test tube and the resin was washed with water to a total volume of 5.2mL filtrate. The title compound was purified by semipreparative reverse-phase HPLC (0.1% trifluoroacetic acid in 0-40% aqueous acetonitrile, C-18 reverse-phase), and lyophilized to afford 14.4g of the title compound in 26% yield. The pure fractions, as analyzed by HPLC, were combined and lyophilized to afford the title compound (2.1mg). MS (M+H=469). The title compound demonstrates specific inhibitory activity toward urokinase. The title compound was shown to have urokinase inhibitory activity in *in vitro* assays.

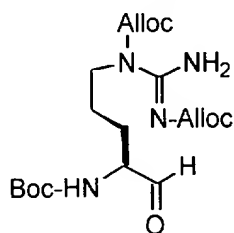
Example 15Preparation of t-butoxycarbonyl-L-Arg(alloc)<sub>2</sub>-ol

To a stirred solution of N<sup>a</sup>-Boc-N<sup>d</sup>-(di-Alloc)-arginine (27.97 g, 63.0 mmol; prepared according to procedure of Loffet and Zhang, 1993, Int. J. Pept. Prot. Res. 42:346-351) and triethylamine (7.65 g, 75.6 mmol, 1.2 equiv.) in anhydrous tetrahydrofuran (50 mL) at -23 °C was added dropwise a solution of isobutyl chloroformate (9.46 g, 69.3 mmol, 1.1 equiv.) in anhydrous tetrahydrofuran (13 mL) for 10 minutes under nitrogen atmosphere. The reaction mixture was warmed to 0 °C for 30 minutes. The white precipitate of triethylammonium chloride was filtered and rinsed with anhydrous tetrahydrofuran (60 mL). The filtrates and the rinses were combined and cooled to -10 °C with stirring under nitrogen atmosphere, and then NaBH<sub>4</sub> (4.29 g, 113 mmol, 1.8 equiv.) was added portionwise in 2 minutes. The reaction mixture was warmed to 0 °C for 30

minutes, and then quenched with water. The reaction mixture was acidified to pH 1 with 1N HCl and the biphasic layers were separated. The aqueous layer was extracted with ethyl acetate (2 x 50 mL). The combined ethyl acetate extracts and tetrahydrofuran layer were washed with brine (2 x 50 mL), and the crude residue was dried over sodium sulfate and filtered. Evaporation of the filtrate and purification by silica gel column chromatography using ethyl acetate/hexane 1/2 afforded the title product (16.24 g, 60.1%) as a colorless oil:  $R_f$  0.20 (methylene chloride/ethyl acetate 4/1);  $^1\text{H}$  (400MHz,  $\text{CDCl}_3$ ): 1.4 (s, 9H), 1.6-1.8 (m, 4H), 3.6 (m, 2H), 3.65 (m, 1H), 4.0 (m, 2H), 4.6 (m, 4H), 5.0 (b, 1H), 5.4 (m, 4H), 6.0 (b, 2H), 9.2 (b, 1H), 9.4 (b, 1H).

### Example 16

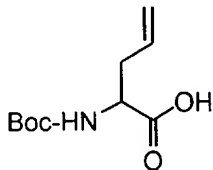
#### Preparation of t-butoxycarbonyl-L-Arg(alloc)<sub>2</sub>-al



To a solution of t-butoxycarbonyl-L-Arg(alloc)<sub>2</sub>-ol from Example 15 (16.24 g, 38.0 mmol) in methylene chloride (190 mL) at ambient temperature were added EDC (36.32 g, 190 mmol, 5 equiv.), dichloroacetic acid (24.5 g, 190 mmol, 5 equiv.), and dimethyl sulfoxide (26 mL). The reaction mixture was stirred for 1 h, and then quenched with water (100 mL). The solvent was removed under reduced pressure. The residue was extracted with diethyl ether (2 x 100 mL). The combined organic phases were washed with a 5% aqueous solution of sodium bicarbonate (250 mL), brine (250 mL), then dried over sodium sulfate and filtered. Evaporation of the filtrate afforded the title product in quantitative yield. The product was used without further purification:  $R_f$  0.57

(hexane/ethyl acetate 1/1);  $^1\text{H}$  (400MHz,  $\text{CDCl}_3$ ): 1.4 (s, 9H), 1.6-1.8 (m, 4H), 4.0 (m, 2H), 4.3 (m, 1H), 4.6 (m, 5H), 5.3 (m, 4H), 5.9-6.0 (m, 3H).



Example 17Preparation of t-butoxycarbonyl-L-Allyl glycine

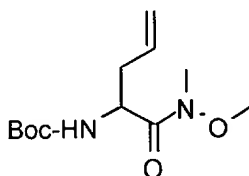
5

To a solution of L-Allyl glycine (25 g, 217.20 mmol, Aldrich) in acetonitrile (350 mL) was added, portion-wise, di-tert-butyl dicarbonate (61.62 g, 282.36 mmol, 1.3 equiv.), while maintaining the reaction at pH 9 to 10 by adding 1N NaOH (282 mL) dropwise.

- 10 After stirring at ambient temperature overnight, the solvent was removed under reduced pressure. The crude residue was partitioned with water and hexane, and the aqueous layer was acidified to pH 3 to 4 with 1N HCl, followed by and extraction 3X with ethyl acetate solvents. The organic phases were washed 1X with 1N HCl, and 1X with H<sub>2</sub>O. The residue was dried over magnesium sulfate and filtered. Evaporation of the filtrate gave
- 15 45.24 g (97% yield) of product as a white solid.  $R_f = 0.34$  (9 methylene chloride/ 1 ethanol). The desired product was judged pure by TLC. <sup>1</sup>H (400 Mhz, CDCl<sub>3</sub>): 1.47 (s, 9H), 2.47-2.76 (b, 2H), 4.40 (b, 1H), 5.23 (b, 1H), 5.15-5.24 (m, 2H), 5.69-5.78 (m, 1H).

Example 18

- 20 Preparation of t-butoxycarbonyl-L-Allyl glycyl-N-methoxy-N-methylamide

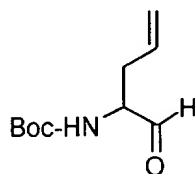


- 4-methylmorpholine (16.92 g, 167.36 mmol, 2.4 equiv.) was added to a solution of t-butoxycarbonyl-L-allyl glycine of Example 17 (15 g, 69.75 mmol), N,O-
- 25 dimethylhydroxylamine hydrochloride (9.52 g, 97.63 mmol, 1.4 equiv.), TBTU (31.35 g,

97.63 mmol, 1.4 equiv.), and 1-hydroxybenzotriazole (13.20 g, 97.63 mmol, 1.4 equiv.) in anhydrous acetonitrile (90 mL). The reaction was stirred at ambient temperature for 16 hours and solvents were removed in vacuo. The residue was partitioned between ethyl acetate and 1N HCl. The aqueous phase was re-extracted with ethyl acetate (3X). The combined organic layers were washed with saturated sodium bicarbonate (1X), and H<sub>2</sub>O (1X) and then dried over magnesium sulfate, filtered, and concentrated in vacuo. The desired product was obtained in 89% mass recovery and was judged pure by TLC.  $R_f$  = 0.37 (1 hexane/ 1 ethyl acetate). <sup>1</sup>H (400 Mhz, CDCl<sub>3</sub>): 1.47 (s, 9H), 2.35-2.56 (bb, 2H), 3.23 (s, 3H), 3.80 (s, 3H), 4.79 (b, 1H), 5.06-5.18 (m, 2H), 5.19-5.24 (b, 1H), 5.71-5.85 (m, 1H).

### Example 19

#### Preparation of t-butoxycarbonyl-L-Allyl glycinal

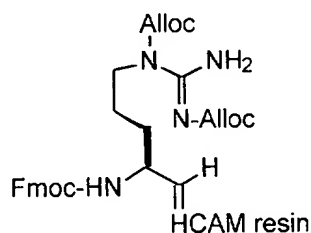


To a solution of t-butoxycarbonyl-L-Allyl glycinal-N-methoxy-N-methylamide of Example 18 (10 g, 38.74 mmol) and anhydrous tetrahydrofuran (58 mL) in a 3 necked round bottomed flask was added dropwise 1M lithium aluminum hydride/tetrahydrofuran (58.12 mL, 1.5 equiv.) at -78°C over 15 minutes under nitrogen atmosphere. The reaction was kept at -78°C with stirring for 1 hr, and then allowed to stir at room temperature for 20 minutes. The reaction mixture was once again cooled to -78°C and quenched slowly with 2M potassium bisulfate (25 mL). The solvent was removed under reduced pressure. The residue was partitioned with ethyl acetate and 1N HCl. The aqueous phase was re-extracted with ethyl acetate (3X), washed with H<sub>2</sub>O (1X), saturated sodium bicarbonate (1X), and H<sub>2</sub>O. The residue was dried over sodium sulfate and filtered. Evaporation of the filtrate afforded product in quantitative yield. The desired product was judged pure by TLC.  $R_f$  = 0.33 (2 hexane/1 ethyl acetate). <sup>1</sup>H (400 Mhz, CDCl<sub>3</sub>): 1.47 (s, 9H), 2.40-

2.66 (bb, 2H), 4.26 (d, 1H), 5.07 (d, 1H), 5.15-5.21 (m, 2H), 5.65-5.80 (m, 1H), 9.60 (s, 1H).

### Example 20

- 5 Preparation of N<sup>a</sup>-Fmoc-N<sup>d</sup>-(di-Alloc)-Arginal-hydrazylcarbonyl-amino methylated polystyrene resin



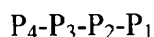
- 10 The title resin was made according to the procedures of Example 15-16 and Example 4, substituting N<sup>a</sup>-Fmoc-N<sup>d</sup>-(di-Alloc)-arginine (Loffet and Zhang, 1993, Int. J. Pept. Prot. Res. 42:346-351) for N<sup>a</sup>-Boc-N<sup>d</sup>-(di-Alloc)-arginine in Example 15.

### Example 21

- 15 Preparation of Libraries of Peptide or Peptidomimetic Compounds - Using the Resin of this Invention in Combinatorial Chemistry:

The invented solid supports also are useful in the field of combinatorial chemistry and development of chemical libraries. With reference to the following formula, it will be

- 20 appreciated that preferred methods within this aspect of the invention result in the development of combinatorial libraries having an aldehyde in the P<sub>1</sub> position, especially argininals and glycinals:



25

According to this aspect of the invention, a library was designed wherein the P<sub>1</sub> residue was kept constant while residues P<sub>4</sub>, P<sub>3</sub>, and P<sub>2</sub> were varied and incorporated into, for

example, peptides and peptidomimetic P<sub>1</sub> aldehydes. Using the invented supports, multiple reactions were carried out in parallel. The P<sub>1</sub> site was first incorporated onto the resin, e.g. the HCAM resin of this invention, (see Examples 1-5), followed by incorporation of variations in the P<sub>4</sub>, P<sub>3</sub>, and P<sub>2</sub> residues, thereby forming a library of peptide or peptidomimetic compounds available for structure-activity analyses in any of a number of *in vitro* or *in vivo* assay systems, including protease inhibition assays.

As will be appreciated by those skilled in the art, this process of library formation and parallel synthesis may be carried out in a number of known formats. In one embodiment, the synthesis was conducted in Whatman mini columns, or the like, wherein standard peptide synthetic methods known in the art were used to extend the peptide chain, with each subsequent coupling being achieved at the carboxy terminus of each added residue. Following synthesis, the multiple peptide variants were cleaved from the resin, isolated, and tested for biological activity, as follows:

Individual members of the library were synthesized in Whatman mini columns for their carboxy terminus using standard peptide synthetic techniques. A total of 20 Whatman mini columns were used. The starting resin containing P<sub>1</sub> aldehyde was used and had the general formula: H<sub>2</sub>N-(CHR)-C=HCAM resin (50 mg resin; substitution: 0.79 mmol/g) which was weighed in each of the 20 mini columns. 1.5 mL of solvent was used in each reaction mini column at each deprotection or coupling reaction, except that 3 X 5 mL of solvent was used during washing steps. The standard Fmoc/tBu/Alloc protection strategy was employed and 1.5 equivalents of coupling reagents were used in each step. All coupling reactions were achieved using PyBOP/DIEA coupling reagents in DMF.

Coupling of the first residue P<sub>2</sub> to P<sub>1</sub> aldehyde was carried out at ambient temperature overnight. The mini columns were drained and washed successively with DMF, DCM, and MeOH or ether. Coupling was determined for each mini column using the quantitative Kaiser ninhydrin test. Double coupling reactions using fresh reagents were carried out when necessary. The resins were then subjected to standard Fmoc cleavage

(using 20% piperidine in DMF, 30 minutes) and another round of acylation was then carried out. The cycle was continued until peptides of the desired length had been assembled.

- 5 The Whatman mini columns were then dried *in vacuo*. The resin in each mini column was treated with a 9:1 mixture of trifluoroacetic acid: water for one hour (1.5 mL). After draining the cleavage products into separate test tubes, the crude products were purified by Sepak column or HPLC and analyzed by mass spec and <sup>1</sup>NMR spectroscopy.

10 Example 22

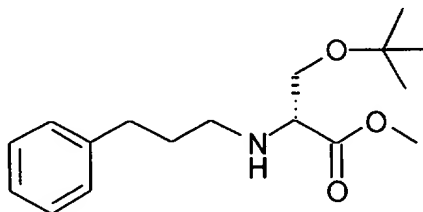
Purification of Peptides or Proteins Using the Resin of this Invention:

- As disclosed herein, the novel resin of this invention provides a convenient platform for manual or automated synthesis of peptides and peptide analogs, including protease  
15 inhibitory transition state analog peptidomimetics. In a further aspect of this invention, due to the facile nucleophilic reaction that occurs between the resin of this invention and aldehyde-bearing molecules, biologically relevant molecules may be purified or isolated by exposure to the resin of this invention under acidic conditions, such that formation of a stable semicarbazone derivative is catalyzed. Subsequent washing steps removes  
20 compounds that do not bear an active aldehyde moiety. Following washing, the free aldehyde may again be released by treatment with strong acid (e.g. TFA treatment). This aspect of the invention is evident from a review of examples 3-8 hereof, wherein an argininal is contacted with a hydrazyl-carbonyl-amino methylated polystyrene resin of this invention, amino acids or amino acid analogs (protected or unprotected) are coupled  
25 to the argininal that has formed a stable semicarbazone with the resin, intermediate deprotection and further coupling steps occur, and finally, the peptide or peptide analog, including a peptidomimetic protease inhibitor is released as the free aldehyde thrombin inhibitor. Likewise for examples 11-14, which exemplify the formation of a urokinase inhibitor. However, it is not required that chemistry be conducted on the immobilized  
30 peptide or protein. In fact, essentially any compound bearing a free aldehyde or a moiety

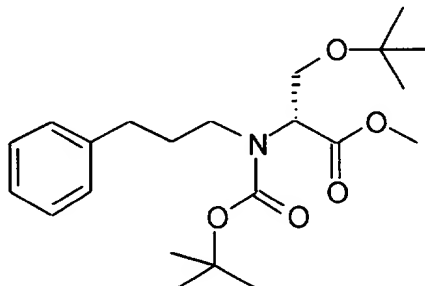
that may be easily converted to an aldehyde (e.g. a free carboxylic acid) may be purified and/or isolated according to this method. By direct analogy, the same procedure may be applied directly to the purification and/or isolation of ketoamide compounds.

### 5 Example 23

#### Preparation of N-alpha-(3-phenylpropyl)-D-serine-t-butyl ether methyl ester



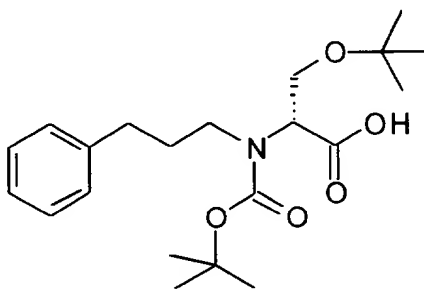
Serine O-t-butyl ether methyl ester (1.50g, 7.1mmol), hydrocinnamaldehyde (1.40 mL, 10.6mmol), and triethylamine (1.18mL, 8.5mmol) were refluxed in tetrahydrofuran (70mL) for 4h. After the solution was allowed to cool to room temperature, sodium borohydride (0.46g, 12mmol) was added in two portions to the stirred solution. The reaction mixture was stirred at ambient temperature for 30 min, and the solution was concentrated under reduced pressure. The residue was partitioned between ethyl acetate 1.0M HCl. The organic layer was washed with 1.0N HCl. The aqueous layer was basified with 40% NaOH to pH 10, then extracted with ethyl acetate (2X). The combined organic layers were dried over sodium sulfate, and the solvent was removed in vacuo. The residue was purified by flash chromatography over silica gel (1x6in column) eluting with 10-30% ethyl acetate/hexanes to afford 150mg (7% yield) of the title compound.  $R_f = 0.60$  (50% ethyl acetate/hexanes).

Example 24Preparation of N-alpha-t-butoxycarbonyl-N-alpha-(3-phenylpropyl)-D-serine-t-butyl ether methyl ester

- 5 The compound of Example 23 (150mg, 0.51mmol), di-t-butyl dicarbonate (167mg, 0.77mmol) and diisopropylethylamine (0.13mL, 0.77mmol) were stirred in tetrahydrofuran (2mL) at ambient temperature overnight. The reaction mixture was concentrated under reduced pressure. The residue was diluted with ethyl acetate (20 mL), washed successively with 1.0N HCl (2X), saturated sodium bicarbonate (2X), and brine (1X), and the solvent was removed in vacuo to afford 206mg of the title compound in
- 10 quantitative yield.  $R_f = 0.74$  (50% ethyl acetate/hexanes).

Example 25Preparation of N-alpha-t-butoxycarbonyl-N-alpha-(3-phenylpropyl)-D-serine-t-butyl ether methyl ester

- 15 methyl ester



- To a solution of the compound of Example 24 (206mg, 0.52mmol) in methanol (3.5mL) was added dropwise 1.0N LiOH (0.63mL, 0.63mmol). The solution became cloudy, then became homogeneous in 5 min. The reaction mixture was allowed to stir at ambient
- 20 temperature overnight. Additional 1.0N LiOH (1.47mL) was added. After 2h, additional 1.0N LiOH (1.0mL) was added. After no starting material was observed by TLC (50%

ethyl acetate/hexanes), the reaction mixture was acidified to pH4 with DOWEX(50X8-400) ion exchange resin. The solution was filtered, rinsing with methanol, then water. The solution was concentrated under reduced pressure, the lyophilized to afford 189mg of the title compound in 95% yield as a yellow oil.  $R_f = 0.04$  (50% ethyl acetate/hexanes).

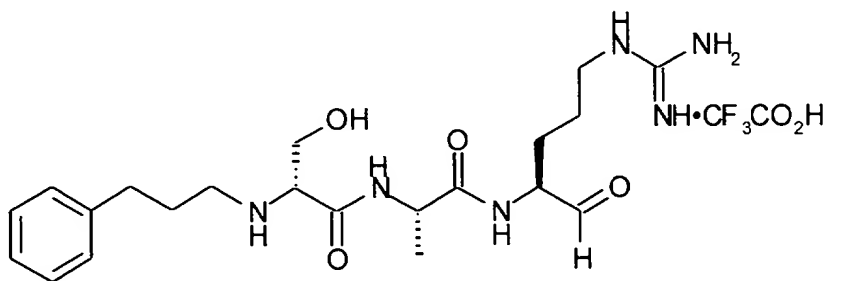
5

Use of the title compound according to step 4 of Example 14 (in place of N-alpha-Fmoc-D-Serine-O-t-butyl ether) results in the production of the Example 26 product, outlined below.

10

#### Example 26

Preparation of N-alpha-(3-phenylpropyl)-D-seryl-alanyl-argininal, trifluoroacetate salt



Steps 1 through 4 and Step 7 of Example 14 were followed, with the substitution in Step 2 of N- $\alpha$ -Fmoc-alanine for N- $\alpha$ -Fmoc-azetidine, and with the substitution in Step 4 of the compound of Example 25 for N- $\alpha$ -Fmoc-D-serine O-t-butyl ether, to make the title compound. The title compound was shown to have urokinase inhibitory activity in *in vitro* assays.

15



**WHAT IS CLAIMED IS:**

- 1           1.       A method for production of a derivatized resin represented by the formula  
2       (I):  
3           (I)        $R_4\text{-NH-(C=X)-Y-Z-SS}$  wherein:  
4                    $R_4$     is  $\text{-NH-R}_3$ ,  $\text{-NH}_2$ ,  $\text{-OH}$ , or  $\text{-O-R}_3$ , wherein  $R_3$  is a protecting  
5                            group, provided that when  $R_4$  is  $\text{-NH-R}_3$  or  $\text{-O-R}_3$ , then the  
6                            protecting group is removed and replaced by  $\text{-H}$  in the final product  
7                            (I);  
8                    $X$     is O, S, or  $\text{NR}_7$ ;  
9                    $R_7$     is H, alkyl, alkenyl, aryl, aralkyl, cycloalkyl, or heterocycle;  
10                   $Y$     is absent,  $\text{-NH-}$ , or  $\text{-CH}_2\text{-}$ ;  
11                   $Z$     is absent or is a substituent selected from the group consisting of  
12                            $\text{-NH-}$ ,  $\text{-O-}$ ,  $\text{-(C=O)-}$ ,  $\text{-S-}$ ,  $\text{SO}_2\text{-}$ , alkyl, alkenyl, aryl, aralkyl,  
13                           cycloalkyl, heterocycle, and combinations thereof, provided that  
14                           when  $Y$  is absent and  $X$  is O or S,  $Z$  does not comprise  $\text{-(C=O)-}$   
15                           immediately adjacent to  $\text{-(C=X)-}$ , and when  $Y$  is  $\text{-NH-}$  and  $Z$   
16                           comprises an  $\text{-NH-}$  or an  $\text{-S-}$ , at least one carbon atom separates  $Y$   
17                           and the  $\text{-NH-}$  or  $\text{-S-}$  of  $Z$ , wherein, under conditions for peptide  
18                           synthesis, functional groups of  $Z$  are protected;  
19                   $SS$     is a solid support;  
20       wherein said derivatized resin represented by formula (I) is prepared by a process  
21       comprising the steps of:  
22       (i)       reacting a starting material represented by formula (C)  
23                   (C)     $R_1\text{-(C=X)-Y-Z-SS}$ , wherein  $R_1$  is a leaving group;  
24                            with a reactant of formula (D)  
25                   (D)     $R_4\text{-NH}_2$   
26                            to form the product (I) of formula  $R_4\text{-NH-(C=X)-Y-Z-SS}$ ; and  
27       (ii)       recovering the derivatized resin (I).

1 2. The method according to claim 1 which further comprises reacting an aldehyde or  
2 ketoamide with the derivatized resin represented by formula (I), thereby producing an  
3 immobilized aldehyde or ketoamide.

1 3. The method according to claim 2, further comprising the step of performing solid-  
2 phase chemistry on said immobilized aldehyde or ketoamide, selected from peptide  
3 synthesis, synthesis of peptide analogs, production of a peptidomimetic compound and  
4 combinatorial chemistry, thereby producing a modified aldehyde or ketoamide, wherein  
5 reactive groups present on a growing peptide or peptide analog chain or peptidomimetic  
6 compound are optionally protected.

1 4. The method according to claim 3, further comprising the step of cleaving,  
2 deprotecting and recovering the modified aldehyde or ketoamide as the free aldehyde or  
3 ketoamide.

1 5. The method according to claim 2, further comprising the step of cleaving,  
2 deprotecting and recovering the aldehyde or ketoamide as the free aldehyde or ketoamide.

1 6. The method according to claim 1, wherein said material represented by formula  
2 (C) is prepared by a process comprising reacting a starting material of formula

3 (A)  $R-Y-Z-SS$ , wherein R is a leaving group,  
4 with a reactant of formula

5 (B)  $R_1-(C=X)-R_2$ , wherein  $R_2$  is a leaving group, same or different  
6 than  $R_1$ ,

7 to form said starting material (C) represented by formula

8 (C)  $R_1-(C=X)-Y-Z-SS$ .

1 7. The method according to claim 6 wherein -R-Y of reactant (A) is  $-NH_2$ , such that  
2 said method produces a derivatized resin represented by formula (IA):

3 (IA)  $R_4-NH-(C=X)-NH-Z-SS$ .

1 8. The method according to claim 6 wherein Y is absent or is -CH<sub>2</sub>-, X is oxygen,  
 2 and R1 is hydroxyl, such that said method produces a derivatized resin represented by  
 3 formula (IB):

4 (IB) R<sub>4</sub>-NH-(C=O)-Y-Z-SS.

1 9. The method according to claim 8 which further comprises thionating the  
 2 derivatized resin (IB) to produce a product represented by formula (IC);

3 (IC) R<sub>4</sub>-NH-(C=S)-Y-Z-SS.

1 10. The method according to claim 9 wherein said thionating comprises contacting  
 2 the product (IB) with Lawesson's reagent or P<sub>2</sub>S<sub>5</sub> in mild base.

1 11. The method according to claim 9 further comprising alkylating the derivatized  
 2 resin (IC) with an alkylating agent capable of contributing an alkyl group R11 to form an  
 3 intermediate represented by the formula (H):

4 (H) R<sub>4</sub>-N=(C-S-R11)-Y-Z-SS, and reacting the intermediate (H) with NH<sub>2</sub>-  
 5 R7 to form a product represented by the formula (ID):

6 (ID) R<sub>4</sub>-NH-(C=NR7)-Y-Z-SS; and recovering the product (ID).

1 12. The method according to claim 11 wherein said alkylating agent is selected from  
 2 the group consisting of iodomethane, iodoethane, methylbromide, ethylbromide,  
 3 allylbromide, allylchloride, dimethylsulfate, and CH<sub>3</sub>OSO<sub>3</sub>CF<sub>3</sub>.

1 13. The method according to claim 1 to produce a product represented by the formula  
 2 (ID), wherein reactant (C), when Y is not absent and R1 is -OH, is prepared by a process  
 3 comprising:

4 reacting a starting material represented by formula (B):

5 (B) R<sub>1</sub>-(C=X)-R<sub>2</sub>, wherein R<sub>1</sub> is -OH and R<sub>2</sub> is an independently selected  
 6 leaving group, same or different than R<sub>1</sub>, and X is NR<sub>7</sub>,

7 with a reactant represented by formula (G):

8 (G) T-Z-SS; wherein T is -CH<sub>2</sub>Cl, -NH<sub>2</sub>, or -COOH, under conditions  
9 permitting reaction of (B) with (C), such that T is transformed into moiety Y, and  
10 recovering the material (C) wherein X is NR<sub>7</sub>.

1 14. The method according to claim 13 wherein the reactant (B) is selected from the  
2 group consisting of diimidazole imine and phosgeneimine diimidazole.

1 15. The method according to claim 1 wherein, when R<sub>4</sub> of the derivatized resin  
2 represented by formula (I) is R<sub>3</sub>-NH, R<sub>4</sub> is converted to a reactive derivatized resin  
3 bearing a free amine by removal of R<sub>3</sub>.

1 16. The method according to claim 15 wherein said reactive derivatized resin is  
2 contacted with an appropriately protected aldehyde or ketoamide to form a semicarbazone  
3 derivatized resin.

1 17. The method according to claim 16 wherein said aldehyde is an argininal having a  
2 guanidino side chain and an amino terminal nitrogen.

1 18. The method according to claim 17 wherein said aldehyde is orthogonally  
2 protected.

1 19. The method according to claim 18 wherein said argininal guanidino side chain is  
2 di-Boc or di-Alloc protected and said amino terminal nitrogen is Fmoc protected.

1 20. The method according to claim 19 further comprising:  
2 (a) deprotecting said amino terminal nitrogen;  
3 (b) linking amino acid residues, amino acid analog residues, peptide or peptide analogs to  
4 said nitrogen to form an immobilized peptide or peptide analog comprising said  
5 aldehyde at the carboxy terminus thereof; and

6 (c) in a single step, deprotecting and cleaving said immobilized peptide or peptide analog  
7 from said resin.

1 21. The method according to claim 20 wherein said single step deprotecting and  
2 cleaving comprises contacting the immobilized peptide or peptide analog and said resin  
3 with trifluoroacetic acid/water for a time between about one hour and about two hours.

1 22. The method according to claim 16 wherein said semicarbazone derivatized resin  
2 is represented by the formula (II):

3 (II)  $\text{PG-NH-C(R}^*)\text{-CH=N-NH-(C=X)-Y-Z-SS,}$

4 wherein (R\*) represents an amino acid side chain and PG represents a protecting group.

1 23. The method according to claim 22 wherein a peptide or peptide analog is  
2 synthesized on said semicarbazone derivatized resin by removal of the protecting group,  
3 PG, and linking amino acids, amino acid analogs, peptide or peptide analogs to the free  
4 amino group revealed upon removal of said protecting group PG, to produce a derivatized  
5 resin represented by formula (III):

6 (III)  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=X)-Y-Z-SS,}$

7 wherein (res)<sub>t</sub> represents a peptide chain of "t" residues, wherein "t" is an integer between  
8 1 and about 50.

1 24. The method according to claim 23 wherein t is an integer between about 3 and 10.

1 25. The method according to claim 23 wherein said (res)<sub>t</sub> is produced by sequential  
2 linkage of monomers or by linkage of one or more oligomeric or polymeric units.

1 26. The method according to claim 23 wherein said derivatized resin represented by  
2 formula (III) is selected from the group consisting of:

3  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=X)-NH-Z-SS;}$

4  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=X)-NH-SS;}$

- 5 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=O)-CH<sub>2</sub>-Z-SS;  
 6 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=O)-Z-SS;  
 7 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=O)-SS;  
 8 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=S)-CH<sub>2</sub>-Z-SS;  
 9 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=S)-Z-SS;  
 10 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=S)-SS;  
 11 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=NR<sub>7</sub>)-CH<sub>2</sub>-Z-SS;  
 12 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=NR<sub>7</sub>)-Z-SS; and  
 13 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=NR<sub>7</sub>)-SS.

1 27. The method according to claim 26 wherein the moiety (res)<sub>t</sub>-NH-C(R\*)-CH=  
 2 represents a resin linked protease inhibitor.

1 28. The method according to claim 2 wherein said reacting is for purpose of purifying  
 2 or isolating an aldehyde- or a ketoamide-containing compound wherein said reacting  
 3 comprises:

4 (i) contacting, under conditions permitting interaction of said aldehyde or  
 5 ketoamide carbonyl in solution with a free amino-terminal amine of a  
 6 derivatized resin of formula:

7 (I) NH<sub>2</sub>-NH-(C=X)-Y-Z-SS; and

8 (b) allowing sufficient time for said interaction to proceed to completion such  
 9 that said aldehyde or ketoamide is bound to said derivatized resin thereby  
 10 removing the aldehyde- or ketoamide-containing compound from the  
 11 solution.

1 29. The method according to claim 3 comprising preparation of a library of peptides  
 2 or peptide analogs comprising the steps of:

3 (i) in each of a series of separate reaction vessels, contacting an aldehyde or  
 4 ketoamide with a derivatized resin represented by formula (I), under conditions which  
 5 permit the formation of a stable semicarbazone between said aldehyde or ketoamide and

6 said resin, thereby producing an immobilized aldehyde or ketoamide as the P<sub>1</sub> residue of a  
 7 nascent peptide or peptide analog;  
 8 (ii) in each of said series of separate reaction vessels, contacting same or  
 9 different amino acid residues or amino acid analog residues with the thus immobilized  
 10 aldehyde or ketoamide under conditions permitting formation of a peptide bond so as to  
 11 produce a series of P<sub>2</sub> residues, same or different, in each of said series of separate  
 12 reaction vessels; and  
 13 (iii) repeating step (ii) as many times as required so as to produce a sequential  
 14 series of P residues to generate a peptide or peptide analog of the desired number of  
 15 residues, with intermediate steps of protection and deprotection of reactive groups present  
 16 on the growing peptide or peptide analog chain, as required.

1 30. The method according to claim 6 wherein R<sub>1</sub> is selected from the group  
 2 consisting of imidazole, p-nitrophenoxy, Cl, succinimidyl, and Me-imidazolium; R<sub>2</sub> is  
 3 selected from the group consisting of imidazole, Cl, succinimidyl, and Me-imidazolium.

1 31. The method according to claim 30 wherein reactant (A) is selected from the group  
 2 consisting of aminomethylated polystyrene resin and 4-methyl benzhydrylamine resin.

1 32. A method for production of a derivatized resin represented by the formula (I):

2 (I) R<sub>4</sub>-NH-(C=X)-Y-Z-SS wherein:

3 R<sub>4</sub> is -NH-R<sub>3</sub>, -NH<sub>2</sub>, -OH, or -O-R<sub>3</sub>, wherein R<sub>3</sub> is a protecting  
 4 group, provided that when R<sub>4</sub> is -NH-R<sub>3</sub> or -O-R<sub>3</sub>, then the  
 5 protecting group is removed and replaced by -H in the final product  
 6 (I);

7 X is O, S, or NR<sub>7</sub>;

8 R<sub>7</sub> is H, alkyl, alkenyl, aryl, aralkyl, cycloalkyl, or heterocycle;

9 Y is absent, -NH-, or -CH<sub>2</sub>-;

10 Z is absent or is a substituent selected from the group consisting of

11 -NH-, -O-, -(C=O)-, -S-, SO<sub>2</sub>-, alkyl, alkenyl, aryl, aralkyl,  
 12 cycloalkyl, heterocycle, and combinations thereof, provided that  
 13 when Y is absent and X is O or S, Z does not comprise -(C=O)-  
 14 immediately adjacent to -(C=X)-, and when Y is -NH- and Z  
 15 comprises an -NH- or an -S-, at least one carbon atom separates Y  
 16 and the -NH- or -S- of Z, wherein, under conditions for peptide  
 17 synthesis, functional groups of Z are protected;

18 SS is a solid support;

19 wherein said derivatized resin represented by formula (I) is prepared by a process  
 20 comprising the steps of:

- 21 (i) reacting a starting material of formula  
 22 (E) R<sub>4</sub>-NH-(C=X)-R<sub>2</sub>, wherein R<sub>2</sub> is a leaving group,  
 23 with a reactant of formula  
 24 (A) R-Y-Z-SS, wherein R is a leaving group,  
 25 to form the derivatized resin (I) of formula R<sub>4</sub>-NH-(C=X)-Y-Z-SS; and  
 26 (iii) recovering the derivatized resin (I).

1 33. The method according to claim 32 which further comprises reacting an aldehyde  
 2 or ketoamide with the derivatized resin represented by formula (I), thereby producing an  
 3 immobilized aldehyde or ketoamide.

1 34. The method according to claim 33, further comprising the step of performing  
 2 solid-phase chemistry on said immobilized aldehyde or ketoamide, selected from peptide  
 3 synthesis, synthesis of peptide analogs, production of a peptidomimetic compound and  
 4 combinatorial chemistry, thereby producing a modified aldehyde or ketoamide, wherein  
 5 reactive groups present on a growing peptide or peptide analog chain or peptidomimetic  
 6 compound are optionally protected.



- 1 35. The method according to claim 34, further comprising the step of cleaving,  
 2 deprotecting and recovering the modified aldehyde or ketoamide as the free aldehyde or  
 3 ketoamide.
- 1 36. The method according to claim 33, further comprising the step of cleaving,  
 2 deprotecting and recovering the aldehyde or ketoamide as the free aldehyde or ketoamide.
- 1 37. The method according to claim 32 wherein said reactant (E) is prepared by a  
 2 process comprising the steps of:
- 3 (i) reacting a starting material of formula  
 4 (B)  $R1-(C=X)-R2$ , wherein R1 and R2 are independently selected  
 5 leaving groups, same or different,  
 6 with a reactant of  
 7 (D)  $R4-NH_2$   
 8 to form said reactant (E) of formula  
 9 (E)  $R4-NH-(C=X)-R2$ ; and recovering the reactant (E).
- 1 38. The method according to claim 32 wherein derivatized resin (I) is prepared by a  
 2 process wherein, when X of reactant (B) is an oxygen, said oxygen in the derivatized  
 3 resin (I)  $R4-NH-(C=O)-Y-Z-SS$  is converted to sulfur by thionation to produce  
 4 derivatized resin (I) of formula  $R4-NH-(C=S)-Y-Z-SS$  and, optionally, alkylating said  
 5 derivatized resin of formula  $R4-NH-(C=S)-Y-Z-SS$  and thereafter contacting with  $NH_2-$   
 6 R7, to produce derivatized resin (I) of formula  $R4-NH-(C=NR7)-Y-Z-SS$ .
- 1 39. The method according to claim 32 wherein, when R4 of the derivatized resin  
 2 represented by formula (I) is  $R3-NH$ , R4 is converted to a reactive derivatized resin  
 3 bearing a free amine by removal of R3.

- 1 40. The method according to claim 39 wherein said reactive derivatized resin is  
 2 contacted with an appropriately protected aldehyde or ketoamide to form a semicarbazone  
 3 derivatized resin.
- 1 41. The method according to claim 40 wherein said aldehyde is an argininal having a  
 2 guanidino side chain and an amino terminal nitrogen.
- 1 42. The method according to claim 41 wherein said aldehyde is orthogonally  
 2 protected.
- 1 43. The method according to claim 42 wherein said argininal guanidino side chain is  
 2 di-Boc or di-Alloc protected and said amino terminal nitrogen is Fmoc protected.
- 1 44. The method according to claim 43 further comprising:  
 2 (a) deprotecting said amino terminal nitrogen;  
 3 (b) linking amino acid residues, amino acid analog residues, peptide or peptide analogs to  
 4 said nitrogen to form an immobilized peptide or peptide analog comprising said  
 5 aldehyde at the carboxy terminus thereof; and  
 6 (c) in a single step, deprotecting and cleaving said immobilized peptide or peptide analog  
 7 from said resin.
- 1 45. The method according to claim 44 wherein said single step deprotecting and  
 2 cleaving comprises contacting the immobilized peptide or peptide analog and said resin  
 3 with trifluoroacetic acid/water for a time between about one hour and about two hours.
- 1 46. The method according to claim 40 wherein said semicarbazone derivatized resin  
 2 is represented by the formula (II):  
 3 (II) PG-NH-C(R\*)-CH=N-NH-(C=X)-Y-Z-SS,  
 4 wherein (R\*) represents an amino acid side chain and PG represents a protecting group.

1 47. The method according to claim 46 wherein a peptide or peptide analog is  
 2 synthesized on said semicarbazone derivatized resin by removal of the protecting group  
 3 PG and linking amino acids, amino acid analogs, peptide or peptide analogs to the free  
 4 amino group revealed upon removal of said protecting group PG, to produce a derivatized  
 5 resin represented by formula (III):

6 (III)  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=X)-Y-Z-SS}$ ,

7 wherein  $(\text{res})_t$  represents a peptide chain of "t" residues, wherein "t" is an integer between  
 8 about 1 and about 50.

1 48. The method according to claim 48 wherein t is an integer between about 3 and 10.

1 49. The method according to claim 46 wherein said  $(\text{res})_t$  is produced by sequential  
 2 linkage of monomers or by linkage of one or more oligomeric or polymeric units.

1 50. The method according to claim 47 wherein said derivatized resin represented by  
 2 formula (III) is selected from the group consisting of:

3  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=X)-NH-Z-SS}$ ;

4  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=X)-NH-SS}$ ;

5  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=O)-CH}_2\text{-Z-SS}$ ;

6  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=O)-Z-SS}$ ;

7  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=O)-SS}$ ;

8  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=S)-CH}_2\text{-Z-SS}$ ;

9  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=S)-Z-SS}$ ;

10  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=S)-SS}$ ;

11  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=NR}_7\text{)-CH}_2\text{-Z-SS}$ ;

12  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=NR}_7\text{)-Z-SS}$ ; and

13  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=NR}_7\text{)-SS}$ .

1 51. The method according to claim 50 wherein  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=}$  represents a  
 2 resin linked protease inhibitor.

1 52. The method according to claim 33 wherein said reacting is for the purpose of  
 2 purifying or isolating an aldehyde- or a ketoamide-containing compound, wherein said  
 3 reacting comprises:

4 (i) contacting, under conditions permitting interaction of said aldehyde or  
 5 ketoamide carbonyl in solution with a free amino-terminal amine of a  
 6 derivatized resin of formula:

7 (I)  $\text{NH}_2\text{-NH-(C=X)-Y-Z-SS}$ ; and

8 (b) allowing sufficient time for said interaction to proceed to completion such  
 9 that said aldehyde or ketoamide is bound to said derivatized resin thereby  
 10 removing the aldehyde- or ketoamide-containing compound from the  
 11 solution.

1 53. The method according to claim 34 comprising preparation of a library of peptides  
 2 or peptide analogs comprising the steps of:

3 (i) in each of a series of separate reaction vessels, contacting an aldehyde or  
 4 ketoamide with a derivatized resin represented by formula (I), under conditions which  
 5 permit the formation of a stable semicarbazone between said aldehyde or ketoamide and  
 6 said resin, thereby producing an immobilized aldehyde or ketoamide as the P<sub>1</sub> residue of a  
 7 nascent peptide or peptide analog;

8 (ii) in each of said series of separate reaction vessels, contacting same or  
 9 different amino acid residues or amino acid analog residues with the thus immobilized  
 10 aldehyde or ketoamide under conditions permitting formation of a peptide bond so as to  
 11 produce a series of P<sub>2</sub> residues, same or different, in each of said series of separate  
 12 reaction vessels; and

13 (iii) repeating step (ii) as many times as required so as to produce a sequential  
 14 series of P residues to generate a peptide or peptide analog of the desired number of  
 15 residues, with intermediate steps of protection and deprotection of reactive groups present  
 16 on the growing peptide or peptide analog chain, as required.

- 1 54. The method according to claim 37 wherein R1 is selected from the group  
 2 consisting of imidazole, p-nitrophenoxy, Cl, succinimidyl, and Me-imidazolium; R2 is  
 3 selected from the group consisting of imidazole, Cl, succinimidyl, and Me-imidazolium.
- 1 55. The method according to claim 32 wherein reactant (A) is selected from the group  
 2 consisting of aminomethylated polystyrene resin and 4-methyl benzhydrylamine resin.
- 1 56. A method for production of a derivatized resin represented by the formula (I):  
 2 (I)  $R_4-NH-(C=X)-Y-Z-SS$   
 3 wherein:  
 4  $R_4$  is  $-NH-R_3$ ,  $-NH_2$ ,  $-OH$ , or  $-O-R_3$ , wherein  $R_3$  is a protecting  
 5 group, provided that when  $R_4$  is  $-NH-R_3$  or  $-O-R_3$ , then the  
 6 protecting group is removed and replaced by  $-H$  in the final product  
 7 (I);  
 8  $X$  is O, S, or  $NR_7$ ;  
 9  $R_7$  is H, alkyl, alkenyl, aryl, aralkyl, cycloalkyl, or heterocycle;  
 10  $Y$  is absent,  $-NH-$ , or  $-CH_2-$ ;  
 11  $Z$  is absent or is a substituent selected from the group consisting of  
 12  $-NH-$ ,  $-O-$ ,  $-(C=O)-$ ,  $-S-$ ,  $SO_2-$ , alkyl, alkenyl, aryl, aralkyl,  
 13 cycloalkyl, heterocycle, and combinations thereof, provided that  
 14 when  $Y$  is absent and  $X$  is O or S,  $Z$  does not comprise  $-(C=O)-$   
 15 immediately adjacent to  $-(C=X)-$ , and when  $Y$  is  $-NH-$  and  $Z$   
 16 comprises an  $-NH-$  or an  $-S-$ , at least one carbon atom separates  $Y$   
 17 and the  $-NH-$  or  $-S-$  of  $Z$ , wherein, under conditions for peptide  
 18 synthesis, functional groups of  $Z$  are protected;  
 19  $SS$  is a solid support;  
 20 said method comprising the steps of:  
 21 (i) reacting a starting material of formula  
 22 (AA)  $R_4-N=C=X$ ,  
 23 with the an anion of formula (BB):

- 24 (BB) (-)Y-Z-SS,  
25 and quenching the reaction with a proton source; and  
26 (ii) recovering the derivatized resin (I).

1 57. The method according to claim 56 wherein said proton source is a strong or weak  
2 acid, selected from the group consisting of water, HCl, and acetic acid.

1 58. The method according to claim 56 wherein the anion Y(-) in reactant (BB) is  
2 generated by treatment of Y with a base selected from the group consisting of: sodium or  
3 potassium hydride, sodium potassium or lithium alkoxide (e.g. ethoxide), potassium tert-  
4 butoxide, lithium diisopropyl amide, and lithium or potassium bis(trimethylsilyl) amide.

1 59. The method according to claim 56 which further comprises reacting an aldehyde  
2 or ketoamide with the derivatized resin represented by formula (I), thereby producing an  
3 immobilized aldehyde or ketoamide.

1 60. The method according to claim 59, further comprising the step of performing  
2 solid-phase chemistry on said immobilized aldehyde or ketoamide, selected from peptide  
3 synthesis, synthesis of peptide analogs, production of a peptidomimetic compound and  
4 combinatorial chemistry, thereby producing a modified aldehyde or ketoamide, wherein  
5 reactive groups present on a growing peptide or peptide analog chain or peptidomimetic  
6 compound are optionally protected.

1 61. The method according to claim 60, further comprising the step of cleaving,  
2 deprotecting and recovering the modified aldehyde or ketoamide as the free aldehyde or  
3 ketoamide.

1 62. The method according to claim 59, further comprising the step of cleaving,  
2 deprotecting and recovering the aldehyde or ketoamide as the free aldehyde or ketoamide.

- 1 63. The method according to claim 56 wherein, when X is oxygen, the reactant (AA)  
2 is an isocyanate, when X is sulfur, the reactant (AA) is thiocyanate, and where X is NR7,  
3 the reactant (AA) is a carbodiimide.
- 1 64. The method according to claim 56 wherein, when R4 of the derivatized resin  
2 represented by formula (I) is R3-NH, R4 is converted to a reactive derivatized resin  
3 bearing a free amine by removal of R3.
- 1 65. The method according to claim 64 wherein said reactive derivatized resin is  
2 contacted with an appropriately protected aldehyde or ketoamide to form a semicarbazone  
3 derivatized resin.
- 1 66. The method according to claim 65 wherein said aldehyde is an argininal having a  
2 guanidino side chain and an amino terminal nitrogen.
- 1 67. The method according to claim 66 wherein said aldehyde is orthogonally  
2 protected.
- 1 68. The method according to claim 67 wherein said argininal guanidino side chain is  
2 di-Boc or di-Alloc protected and said amino terminal nitrogen is Fmoc protected.
- 1 69. The method according to claim 68 further comprising:  
2 (a) deprotecting said amino terminal nitrogen;  
3 (b) linking amino acid residues, amino acid analog residues, peptide or peptide analogs to  
4 said nitrogen to form an immobilized peptide or peptide analog comprising said  
5 aldehyde at the carboxy terminus thereof; and  
6 (c) in a single step, deprotecting and cleaving said immobilized peptide or peptide analog  
7 from said resin.

1 70. The method according to claim 69, wherein said single step deprotecting and  
 2 cleaving comprises contacting the immobilized peptide or peptide analog and said resin  
 3 with trifluoroacetic acid/water for a time between about one hour and about two hours.

1 71. The method according to claim 65 wherein said semicarbazone derivatized resin  
 2 is represented by the formula (II):

3 (II)  $\text{PG-NH-C(R}^*)\text{-CH=N-NH-(C=X)-Y-Z-SS}$ ,

4 wherein (R<sup>\*</sup>) represents an amino acid side chain and PG represents a protecting group.

1 72. The method according to claim 70 wherein a peptide or peptide analog is  
 2 synthesized on said semicarbazone derivatized resin by removal of the protecting group  
 3 PG and linking amino acids, amino acid analogs, peptide or peptide analogs to the free  
 4 amino group revealed upon removal of said protecting group PG, to produce a derivatized  
 5 resin represented by formula (III):

6 (III)  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=X)-Y-Z-SS}$ ,

7 wherein (res)<sub>t</sub> represents a peptide chain of "t" residues, wherein "t" is an integer between  
 8 1 and about 50.

1 73. The method according to claim 72 wherein t is an integer between about 3 and 10.

1 74. The method according to claim 72 wherein said (res)<sub>t</sub> is produced by sequential  
 2 linkage of monomers or by linkage of one or more oligomeric or polymeric units.

1 75. The method according to claim 72 wherein said derivatized resin represented by  
 2 formula (III) is selected from the group consisting of:

3  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=X)-NH-Z-SS}$ ;

4  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=X)-NH-SS}$ ;

5  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=O)-CH}_2\text{-Z-SS}$ ;

6  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=O)-Z-SS}$ ;

7  $(\text{res})_t\text{-NH-C(R}^*)\text{-CH=N-NH-(C=O)-SS}$ ;



8 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=S)-CH<sub>2</sub>-Z-SS;  
 9 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=S)-Z-SS;  
 10 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=S)-SS;  
 11 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=NR<sub>7</sub>)-CH<sub>2</sub>-Z-SS;  
 12 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=NR<sub>7</sub>)-Z-SS; and  
 13 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=NR<sub>7</sub>)-SS.

1 76. The method according to claim 75 wherein (res)<sub>t</sub>-NH-C(R\*)-CH= represents a  
 2 resin-bound protease inhibitor.

1 77. The method according to claim 59 wherein said reacting is for purpose of  
 2 purifying or isolating an aldehyde- or a ketoamide-containing compound, wherein said  
 3 reacting comprises:

4 (i) contacting, under conditions permitting interaction of said aldehyde or  
 5 ketoamide carbonyl in solution with a free amino-terminal amine of a  
 6 derivatized resin of formula:

7 (I) NH<sub>2</sub>-NH-(C=X)-Y-Z-SS; and

8 (b) allowing sufficient time for said interaction to proceed to completion such  
 9 that said aldehyde or ketoamide is bound to said derivatized resin thereby  
 10 removing the aldehyde- or ketoamide-containing compound from the  
 11 solution.

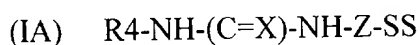
1 78. The method according to claim 60 comprising preparation of a library of peptides  
 2 or peptide analogs comprising the steps of:

3 (i) in each of a series of separate reaction vessels, contacting an aldehyde or  
 4 ketoamide with a derivatized resin represented by formula (I), under conditions which  
 5 permit the formation of a stable semicarbazone between said aldehyde or ketoamide and  
 6 said resin, thereby producing an immobilized aldehyde or ketoamide as the P<sub>1</sub> residue of a  
 7 nascent peptide or peptide analog;

(ii) in each of said series of separate reaction vessels, contacting same or different amino acid residues or amino acid analog residues with the thus immobilized aldehyde or ketoamide under conditions permitting formation of a peptide bond so as to produce a series of P<sub>2</sub> residues, same or different, in each of said series of separate reaction vessels; and

(iii) repeating step (ii) as many times as required so as to produce a sequential series of P residues to generate a peptide or peptide analog of the desired number of residues, with intermediate steps of protection and deprotection of reactive groups present on the growing peptide or peptide analog chain, as required.

79. A method for production of a derivatized resin represented by the formula (IA):



wherein:

R<sub>4</sub> is -NH-R<sub>3</sub>, -NH<sub>2</sub>, -OH, or -O-R<sub>3</sub>, wherein R<sub>3</sub> is a protecting group, provided that when R<sub>4</sub> is -NH-R<sub>3</sub> or -O-R<sub>3</sub>, then the protecting group is removed and replaced by -H in the final product (IA);

X is O, S, or NR<sub>7</sub>;

R<sub>7</sub> is H, alkyl, alkenyl, aryl, aralkyl, cycloalkyl, or heterocycle;

Z is absent or is a substituent selected from the group consisting of -NH-, -O-, -(C=O)-, -S-, SO<sub>2</sub>-, alkyl, alkenyl, aryl, aralkyl, cycloalkyl, heterocycle, and combinations thereof, provided that when Z comprises an -NH- or an -S-, at least one carbon atom separates said -NH- or -S-, and the -NH- immediately adjacent to the -(C=X)-, wherein, under conditions for peptide synthesis, functional groups of Z are protected; and

SS is a solid support;

said method comprising the steps of:

(i) reacting a starting material of formula :



21 with a reactant of formula:

22 (F)  $R_4-NH-(C=X)-NH-Q$ ;

23 to form the derivatized resin (IA);

24 (ii) recovering the derivatized resin (IA);

25 wherein:

26 T is  $-CH_2Cl$ ,  $-NH_2$ , or  $-COOH$ , and

27 Q is  $-R_9-R_{10}$ , wherein  $R_9$  is substituted or unsubstituted, alkyl, alkenyl, aryl, aralkyl,

28 cycloalkyl and heterocycle, and  $R_{10}$  is a functional group selected from the group

29 consisting of  $-NH_2$ ,  $-OH$ ,  $-COOH$ ,  $-COCl$ ,  $-SH$ ,  $-SO_2Cl$ ,  $-SO_2H$ ,  $-SO_3$ -lower alkyl,

30 provided that the terminal hydrogen, halogen or a reactive moiety of  $R_{10}$  may be replaced

31 or protected by a protecting group,  $-R_8$ , and moiety Q is selected such that the terminal

32 substituent thereof is reactive with the terminal substituent of T within reactant (G),

33 permitting bond formation between (G) and (F), upon contact thereof.

34

1 80. The method according to claim 79 which further comprises reacting an aldehyde  
2 or ketoamide with the derivatized resin represented by formula (I), thereby producing an  
3 immobilized aldehyde or ketoamide.

1 81. The method according to claim 80, further comprising the step of performing  
2 solid-phase chemistry on said immobilized aldehyde or ketoamide, selected from peptide  
3 synthesis, synthesis of peptide analogs, production of a peptidomimetic compound and  
4 combinatorial chemistry, thereby producing a modified aldehyde or ketoamide, wherein  
5 reactive groups present on a growing peptide or peptide analog chain or peptidomimetic  
6 compound are optionally protected.

1 82. The method according to claim 81, further comprising the step of cleaving,  
2 deprotecting and recovering the modified aldehyde or ketoamide as the free aldehyde or  
3 ketoamide.

1 83. The method according to claim 80, further comprising the step of cleaving,  
2 deprotecting and recovering the aldehyde or ketoamide as the free aldehyde or ketoamide.

1 84. The method according to claim 79 wherein reactant (F) of formula:

2 (F)  $R_4-NH-(C=X)-NH-Q$ ;

3 is prepared by a process comprising:

4 (i) reacting a starting material (N) of formula:

5 (N)  $H_2N-Q-R_8$ ,

6 with a reactant (B) of formula:

7 (B)  $R_1-(C=X)-R_2$ , wherein  $R_1$  and  $R_2$  are independently selected

8 leaving groups, same or different, to form an intermediate (P) of formula:

9 (P)  $R_1-(C=X)-NH-Q-R_8$ ;

10 (ii) reacting intermediate (P) with reactant (D) of formula

11 (D)  $R_4-NH_2$

12 to form an intermediate (U) of formula:

13 (U)  $R_4-NH-(C=X)-NH-Q-R_8$ ; and

14 (iii) removing  $R_8$  to form reactant (F), and recovering reactant (F).

1 85. The method according to claim 79 wherein X is oxygen, and the derivatized resin  
2 is represented by formula (IE):

3 (IE)  $R_4-NH-(C=O)-NH-Z-SS$

4 comprising the steps of:

5 (i) reacting a starting material of formula (W):

6 (W)  $HOOC-NH-Q-R_8$

7 with reactant (D) of formula

8 (D)  $R_4-NH_2$

9 to form intermediate (J) of formula:

10 (J)  $R_4-NH-(C=O)-NH-Q-R_8$ ;

11 (ii) removing  $R_8$  from intermediate (J) to form intermediate (K) of formula:

12 (K)  $R_4-NH-(C=O)-NH-Q$ ; and

- 13 (iii) reacting the intermediate (K) with starting material (G) of formula:  
 14 (G) T-Z-SS to form the derivatized resin (IE) of formula:  
 15 (IE) R4-NH-(C=O)-NH-Z-SS.

1 86. The method according to claim 85 for preparing a derivatized resin represented by  
 2 formula (IF):

3 (IF) R4-NH-(C=S)-NH-Z-SS

4 said method comprising, prior to removal of R8 from intermediate (J):

5 (i) thionating intermediate (J) to form intermediate (L) of formula:

6 (L) R4-NH-(C=S)-NH-Q-R8;

7 (ii) removing R8 from intermediate (L); and

8 (iii) reacting deprotected reactant (L) with reactant (G) T-Z-SS to form derivatized  
 9 resin (IF) wherein X is sulfur.

1 87. The method according to claim 86 for preparing a derivatized resin represented by  
 2 formula (IG):

3 (IG) R4-NH-(C=NR7)-NH-Z-SS

4 said method comprising the step of, prior to deprotection of intermediate (L) in step (ii) of  
 5 claim 86:

6 (i) contacting reactant (L) with an alkylating agent capable of contributing an  
 7 alkyl group R11 to form intermediate (M) of formula:

8 (M) R4-N=(C-S-R11)-Q-R8;

9 (ii) reacting intermediate (M) with reactant (H) of formula:

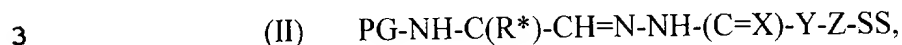
10 (H) NH<sub>2</sub>-R7;

11 (iii) deprotecting and contacting the product with T-Z-SS (G) to form the  
 12 derivatized resin of formula (IG), wherein X is NR7.

1 88. The method according to claim 87 wherein said alkylating agent is selected from  
 2 the group consisting of iodomethane, iodoethane, methylbromide, ethylbromide,  
 3 allylbromide, allylchloride, dimethylsulfate, and CH<sub>3</sub>OSO<sub>2</sub>CF<sub>3</sub>.

- 1 89. The method according to claim 79 wherein, when R4 of the derivatized resin  
2 represented by formula (I) is R3-NH, R4 is converted to a reactive derivatized resin  
3 bearing a free amine by removal of R3.
- 1 90. The method according to claim 89 wherein said reactive derivatized resin is  
2 contacted with an appropriately protected aldehyde or ketoamide to form a semicarbazone  
3 derivatized resin.
- 1 91. The method according to claim 90 wherein said aldehyde is an argininal having a  
2 guanidino side chain and an amino terminal nitrogen.
- 1 92. The method according to claim 91 wherein said aldehyde is orthogonally  
2 protected.
- 1 93. The method according to claim 92 wherein said argininal guanidino side chain is  
2 di-Boc or di-Alloc protected and said amino terminal nitrogen is Fmoc protected.
- 1 94. The method according to claim 93 further comprising:  
2 (a) deprotecting said amino terminal nitrogen;  
3 (b) linking amino acid residues, amino acid analog residues, peptide or peptide analogs  
4 are to said nitrogen to form an immobilized peptide or peptide analog comprising said  
5 aldehyde at the carboxy terminus thereof; and  
6 (c) in a single step, deprotecting and cleaving said immobilized peptide or peptide analog  
7 from said resin.
- 1 95. The method according to claim 94 wherein said single step deprotecting and  
2 cleaving comprises contacting the immobilized peptide or peptide analog and said resin  
3 with trifluoroacetic acid/water for a time between about one hour and about two hours.

1 96. The method according to claim 90 wherein said semicarbazone derivatized resin  
2 is represented by the formula (II):



4 wherein (R<sup>\*</sup>) represents an amino acid side chain and PG represents a protecting group.

1 97. The method according to claim 96 wherein a peptide or peptide analog is  
2 synthesized on said semicarbazone derivatized resin by removal of the protecting group  
3 PG and linking amino acids, amino acid analogs, peptide or peptide analogs to the free  
4 amino group revealed upon removal of said protecting group PG, to produce a derivatized  
5 resin represented by formula (III):

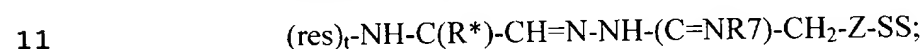
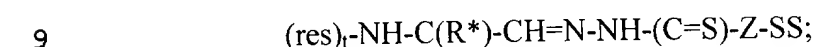
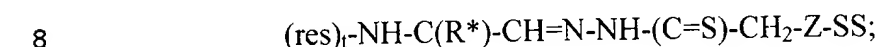
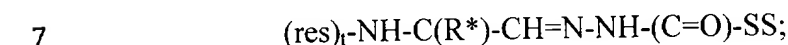
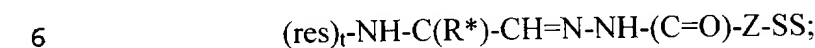
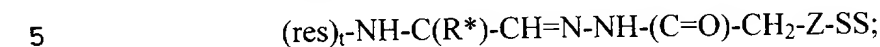
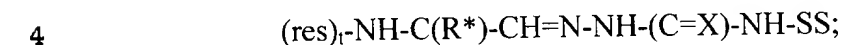


7 wherein (res)<sub>t</sub> represents a peptide chain of "t" residues, wherein "t" is an integer between  
8 about 1 and about 50.

1 98. The method according to claim 97 wherein t is an integer between about 3 and 10.

1 99. The method according to claim 98 wherein said (res)<sub>t</sub> is produced by sequential  
2 linkage of monomers or by linkage of one or more oligomeric or polymeric units.

1 100. The method according to claim 97 wherein said derivatized resin represented by  
2 formula (III) is selected from the group consisting of:



- 12 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=NR7)-Z-SS; and  
 13 (res)<sub>t</sub>-NH-C(R\*)-CH=N-NH-(C=NR7)-SS.

1 101. The method according to claim 100 wherein (res)<sub>t</sub>-NH-C(R\*)-CH= represents a  
 2 resin-bound protease inhibitor.

1 102. The method according to claim 80 wherein said reacting is for purpose of  
 2 purifying or isolating an aldehyde- or a ketoamide-containing compound, wherein said  
 3 reacting comprises:

4 (i) contacting, under conditions permitting interaction of said aldehyde or  
 5 ketoamide carbonyl in solution with a free amino-terminal amine of a  
 6 derivatized resin of formula:

7 (I) NH<sub>2</sub>-NH-(C=X)-Y-Z-SS; and

8 (b) allowing sufficient time for said interaction to proceed to completion such  
 9 that said aldehyde or ketoamide is bound to said derivatized resin thereby  
 10 removing the aldehyde- or ketoamide-containing compound from the  
 11 solution.

1 103. The method according to claim 81 comprising preparation of a library of peptides  
 2 or peptide analogs comprising the steps of:

3 (i) in each of a series of separate reaction vessels, contacting an aldehyde or  
 4 ketoamide with a derivatized resin represented by formula (I), under conditions which  
 5 permit the formation of a stable semicarbazone between said aldehyde or ketoamide and  
 6 said resin, thereby producing an immobilized aldehyde or ketoamide as the P<sub>1</sub> residue of a  
 7 nascent peptide or peptide analog;

8 (ii) in each of said series of separate reaction vessels, contacting same or  
 9 different amino acid residues or amino acid analog residues with the thus immobilized  
 10 aldehyde or ketoamide under conditions permitting formation of a peptide bond so as to  
 11 produce a series of P<sub>2</sub> residues, same or different, in each of said series of separate  
 12 reaction vessels; and



13 (iii) repeating step (ii) as many times as required so as to produce a sequential  
14 series of P residues to generate a peptide or peptide analog of the desired number of  
15 residues, with intermediate steps of protection and deprotection of reactive groups present  
16 on the growing peptide or peptide analog chain, as required.

1 104. The method according to claim 84 wherein R1 is selected from the group  
2 consisting of imidazole, p-nitrophenoxy, Cl, succinimidyl, and Me-imidazolium; R2 is  
3 selected from the group consisting of imidazole, Cl, succinimidyl, and Me-imidazolium.

1 105. The method according to claim 79 wherein reactant (G) is selected from the group  
2 consisting of aminomethylated polystyrene resin and 4-methyl benzhydrylamine resin.

1 106. A method for preparing a derivatized resin which comprises:  
2 (i) contacting an amino methylated polystyrene resin with 1,1-carbonyldiimidazole in  
3 dimethylformamide;  
4 (ii) contacting the product of step (i) with hydrazine in N,N-dimethylformamide; and  
5 (iii) recovering the deprotected, derivatized resin.

1 107. The method according to claim 106, further comprising contacting the derivatized  
2 resin with a di-Boc side-chain-protected, Fmoc amino-protected argininal, or a di-Alloc  
3 side-chain-protected, Boc amino-protected argininal to produce a protected immobilized  
4 argininal.

1 108. The method according to claim 107, further comprising conducting solid phase  
2 peptide synthesis on the protected immobilized argininal, by removing the amino  
3 protection and reaction of activated amino acids or peptides with the deprotected amino  
4 group of the immobilized argininal to produce an immobilized peptide comprising an  
5 argininal at the carboxy terminus thereof.

1 109. The method according to claim 108, further comprising, in a single step,  
2 deprotecting and cleaving the immobilized peptide from the resin, thereby releasing the  
3 immobilized peptide comprising an argininal at the carboxy terminus thereof.

1 110. The method according to claim 109 wherein said single step deprotecting and  
2 cleaving comprises contacting the immobilized peptide or peptide analog and said resin  
3 with trifluoroacetic acid/water for a time between about one hour and about two hours.

1 111. A method for preparing a derivatized resin which comprises:  
2 (i) contacting 1,1-carbonyldiimidazole in dimethylformamide with t-butylcarbazate;  
3 (ii) contacting the product of step (i) with amino methylated polystyrene resin under an  
4 atmosphere of nitrogen;  
5 (iii) removing the t-butyl protection by treatment with trifluoroacetic acid in  
6 dichloromethane in the presence of thioanisole under a nitrogen atmosphere; and  
7 (iv) recovering the deprotected, derivatized resin.

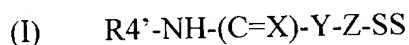
1 112. The method according to claim 111, further comprising contacting the derivatized  
2 resin with a di-Boc side-chain-protected, Fmoc amino-protected argininal, or a di-Alloc  
3 side-chain-protected, Boc amino-protected argininal to produce a protected immobilized  
4 argininal.

1 113. The method according to claim 112, further comprising conducting solid phase  
2 peptide synthesis on the protected immobilized argininal, by removing the amino  
3 protection and reaction of activated amino acids or peptides with the deprotected amino  
4 group of the immobilized argininal to produce an immobilized peptide comprising an  
5 argininal at the carboxy terminus thereof.

1 114. The method according to claim 113, further comprising, in a single step,  
2 deprotecting and cleaving the immobilized peptide from the resin, thereby releasing the  
3 immobilized peptide comprising an argininal at the carboxy terminus thereof.

1 115. The method according to claim 114 wherein said single step deprotecting and  
 2 cleaving comprises contacting the immobilized peptide or peptide analog and said resin  
 3 with trifluoroacetic acid/water for a time between about one hour and about two hours.

1 116. A method for production of an hydroxamic ester, which comprises preparing a  
 2 solid support of formula:



4 wherein:

5  $R4'$  is -OH, or -OR<sub>3</sub>, wherein R<sub>3</sub> is a protecting group, provided that  
 6 when  $R4'$  is -OR<sub>3</sub>, then the protecting group is removed and  
 7 replaced by -H in the final derivatized resin (I);

8 X is O, S, or NR<sub>7</sub>;

9 R<sub>7</sub> is H, alkyl, alkenyl, aryl, aralkyl, cycloalkyl, or heterocycle;

10 Y absent, -NH-, -CH<sub>2</sub>-;

11 Z is absent or is a substituent selected from the group consisting of  
 12 -NH-, -O-, -(C=O)-, -S-, SO<sub>2</sub>-, alkyl, alkenyl, aryl, aralkyl,  
 13 cycloalkyl, heterocycle, and combinations thereof, provided that  
 14 when Y is -NH-, if Z comprises an -NH- or a -S-, at least one  
 15 carbon atom separates Y and Z;

16 SS is a solid support;

17 said method comprising the steps of:

18 (i) reacting a starting material of formula:



20 leaving groups, same or different, with a reactant of formula:



22 to form the derivatized resin (I);

23 (ii) recovering the derivatized resin (I); and

24 (iii) esterifying the resin at the free amino hydroxyl group.

**ABSTRACT OF THE DISCLOSURE**

This invention provides a method for producing a derivatized resin useful in the arts of solid-phase peptide synthesis, combinatorial chemistry, and peptide or protein purification and separation. Methods for synthesizing the derivatized resin, the prototypical example of which is hydrazyl-carbonyl-amino methylated polystyrene, are provided by this invention disclosure.

## DECLARATION (37 CFR 1.63) AND POWER OF ATTORNEY

As below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name, and

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled **RESIN DERIVATIZATION METHOD AND USES THEREOF**, the specification for which

X is attached hereto.  
was filed \_\_\_\_\_, Serial No. \_\_\_\_\_.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment specifically referred to in the oath or declaration.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, Section 1.56(a).

I hereby claim the benefit under Title 35, United States Code, '120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application(s) in the manner provided by the first paragraph of Title 35, United States Code, '112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, '1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

Application Serial No.	Filing Date	Status (Patented, Pending, Abandoned)
---------------------------	-------------	--

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

I hereby appoint the following persons registered to practice before the Patent and Trademark Office as my attorneys with full power of substitution and revocation to prosecute this application and all divisions and continuations thereof and to transact all business in the Patent and Trademark Office connected therewith: Gerard H. Bencen, Reg. No. 35,746.

I request that all correspondence be sent to:

Gerard H. Bencen  
426 Anderson Court  
Orlando, Florida 32801

I further request that all telephone communications be directed to:

Gerard H. Bencen  
407-246-0444

0916576 "072498



IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Examiner : Unknown  
Art Unit : Unknown  
Applicant(s) : Siev, Daniel V., et al.  
Serial No. : Unknown  
Filing Date : July 24, 1998  
For : RESIN DERIVATIZATION METHOD AND USES  
THEREOF

Assistant Commissioner of Patents and Trademarks  
Washington, D.C. 20231

CERTIFICATE OF MAILING BY EXPRESS MAIL (37 CFR 1.10)

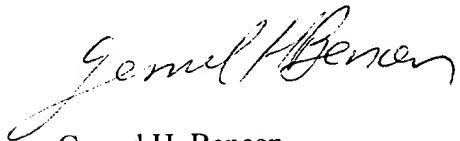
Sir:

Enclosed herewith are the following documents in reference to a new patent application, Docket No. CVS-1:

1. Application (specification and claims) - 90 pages.
2. Fee Transmittal form and Fee (\$3230.00).
3. Utility Application Transmittal Letter.
4. Unexecuted Declaration and Power of Attorney of the Inventors.
5. Return Postcard evidencing receipt of these materials.

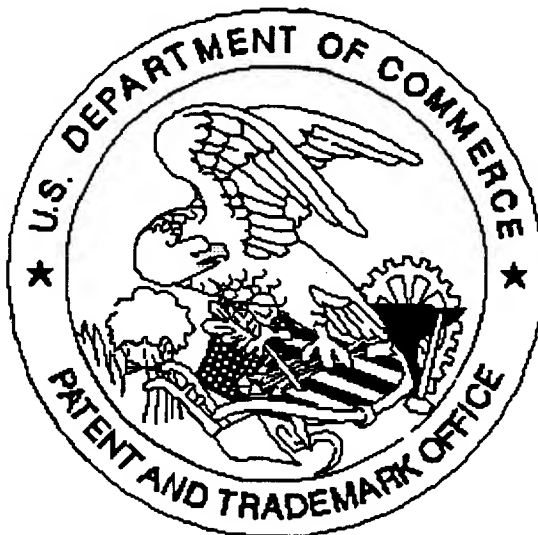
I hereby certify that this correspondence is being deposited with the United States Postal Services "Express Mail Post Office to Addressee" service, under 37 CFR 1.10, and is addressed to the Assistant Commissioner of Patents and Trademarks, Washington, D.C. 20231 on July 15, 1998. Express Mail No. : EE470986382US

Respectfully submitted,



Gerard H. Bencen  
Patent Attorney  
Registration No. 35,746  
Phone No.: 407-246-0444  
Address : 426 Anderson Court  
Orlando, Florida 32801

United States Patent & Trademark Office  
Office of Initial Patent Examination -- Scanning Division



Application deficiencies found during scanning:

1. Application papers are not suitable for scanning and are not in compliance with 37 CFR 1.52 because:
  - ☐ All sheets must be the same size and either A4 (21 cm x 29.7 cm) or 8-1/2" x 11". Pages \_\_\_\_\_ do not meet these requirements.
  - ☐ Papers are not flexible, strong, smooth, non-shiny, durable, and white.
  - ☐ Papers are not typewritten or mechanically printed in permanent ink on one side.
  - ☐ Papers contain improper margins. Each sheet must have a left margin of at least 2.5 cm (1") and top, bottom and right margins of at least 2.0 cm (3/4").
  - ☐ Papers contain hand lettering.
2. Drawings are not in compliance and were not scanned because:
  - ☐ The drawings or copy of drawings are not suitable for electronic reproduction.
  - ☐ All drawings sheets are not the same size. Pages must be either A4 (21 cm x 29.7 cm) or 8-1/2" x 11".
  - ☐ Each sheet must include a top and left margin of at least 2.5 cm (1"), a right margin of at least 1.5 cm (9/16") and a bottom margin of at least 1.0 cm (3/8").
3. Page(s) \_\_\_\_\_ are not of sufficient clarity, contrast and quality for electronic reproduction.
4. Page(s) \_\_\_\_\_ are missing.
5. OTHER: 110 1 Drawing